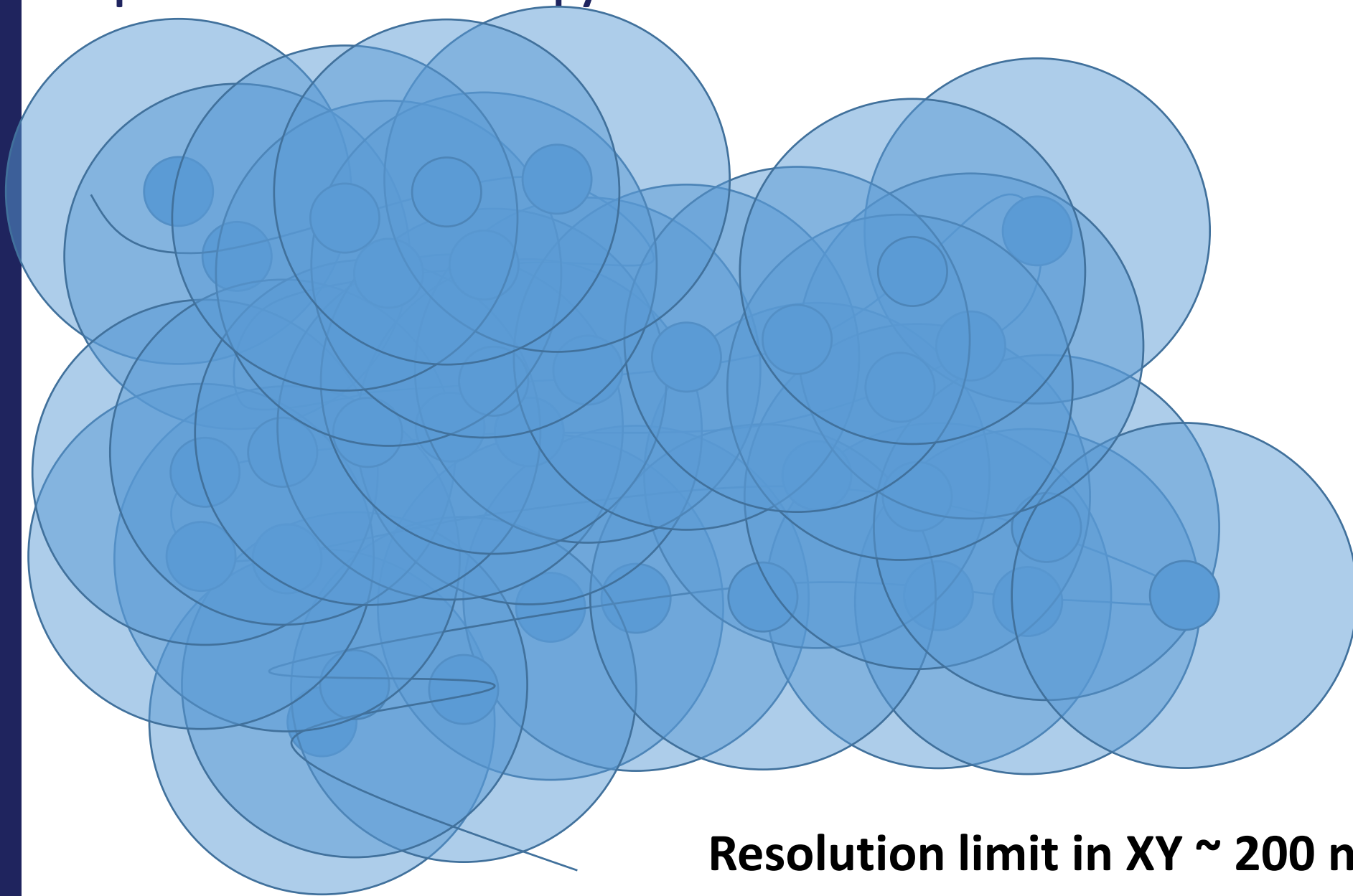


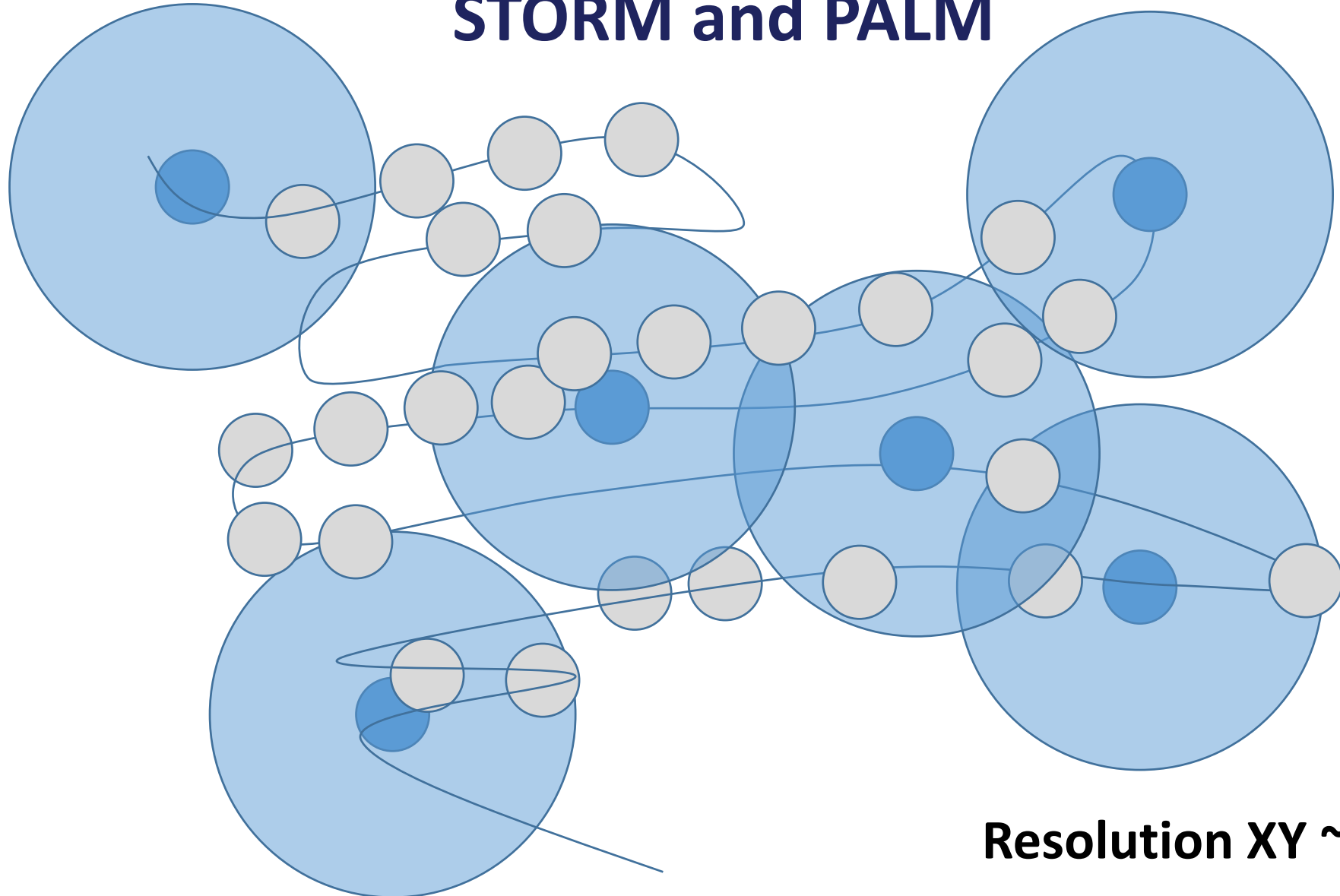
## Superresolution microscopy



**Resolution limit in XY  $\sim$  200 nm**

Superresolution microscopy – single molecule localization microscopy

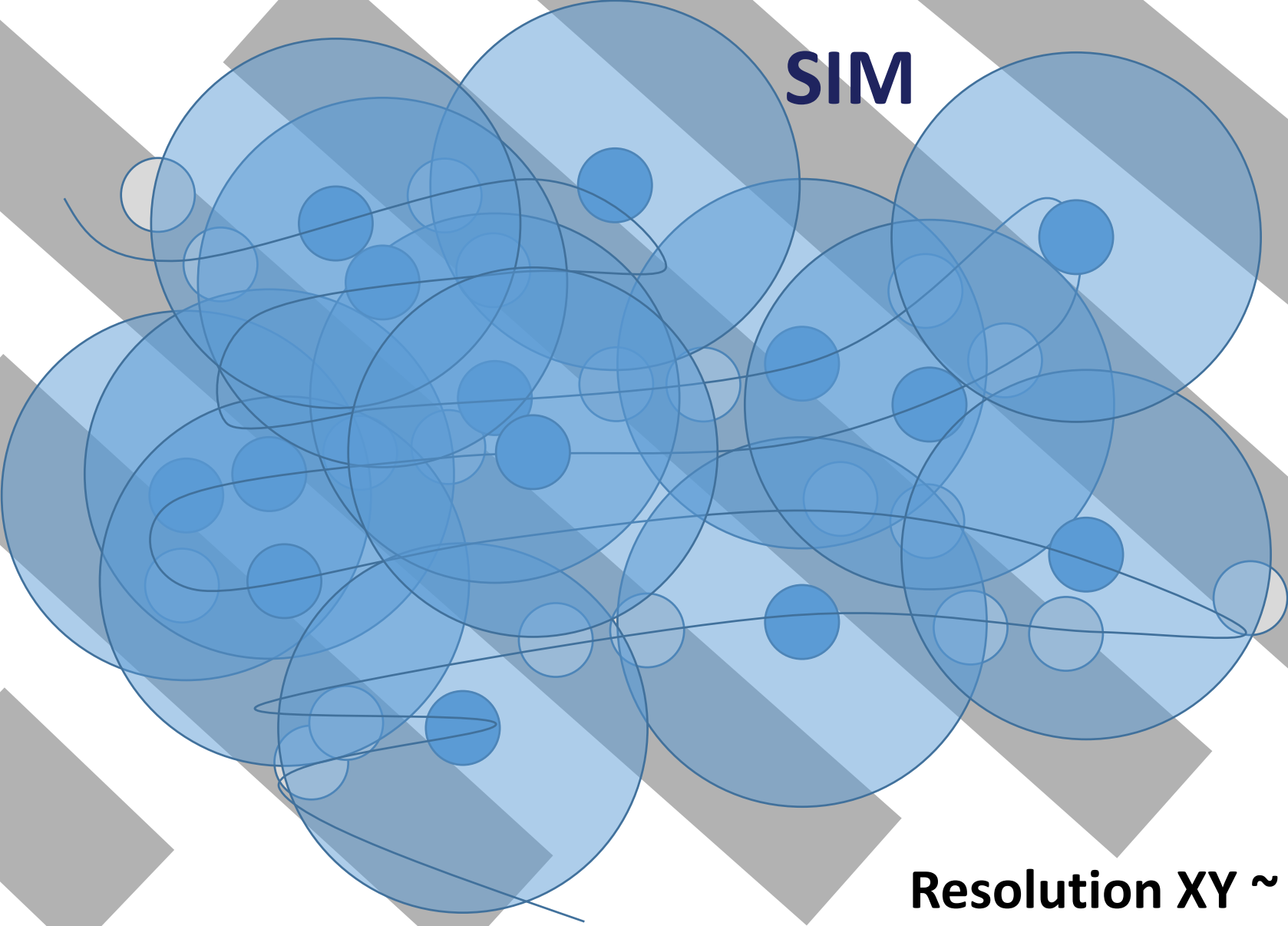
## STORM and PALM



Resolution XY ~ 25 nm

**Superresolution microscopy – Structured illumination**

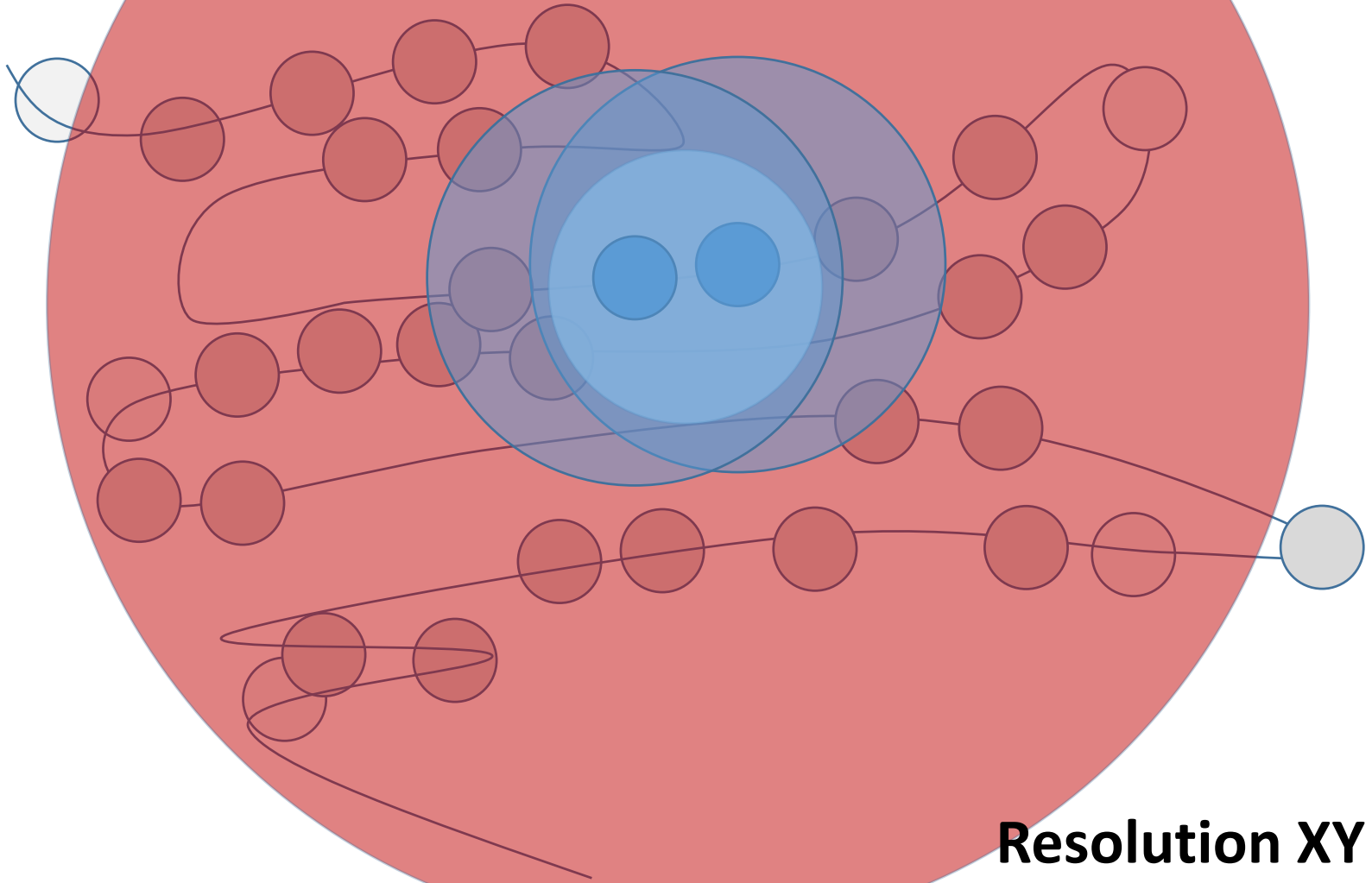
**SIM**



**Resolution XY ~ 120 nm**

**Superresolution microscopy – stimulated emission depletion**

**STED**



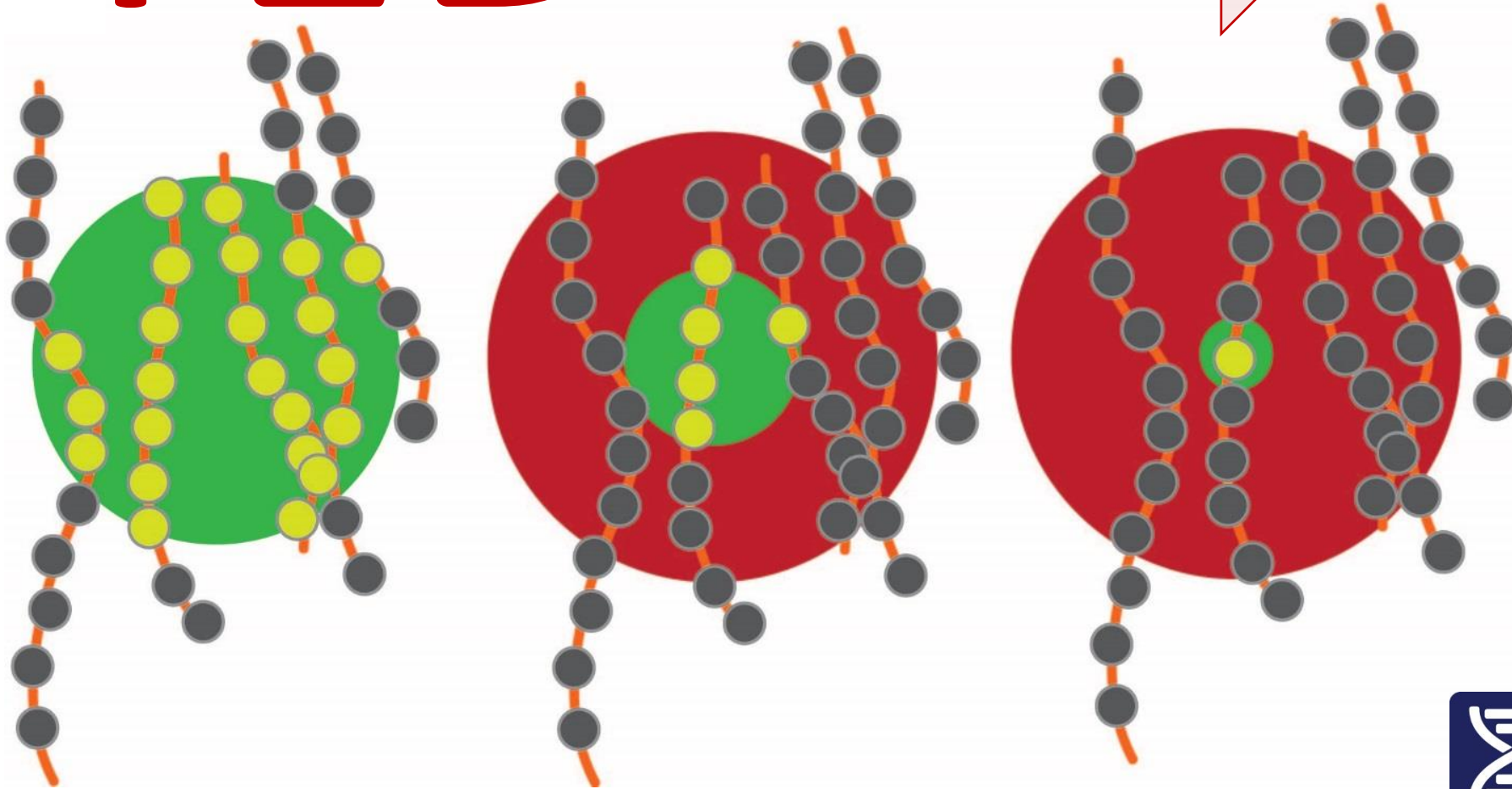
**Resolution XY ~ 40 nm**

# STED

$$d = \frac{\lambda}{2NA}$$

$$d_{STED} = \frac{\lambda}{2NA\sqrt{1 + \xi}}$$

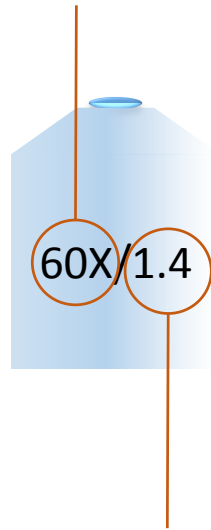
break through the resolution barrier



## Magnification

“How many times the apparent size of the object is increased”

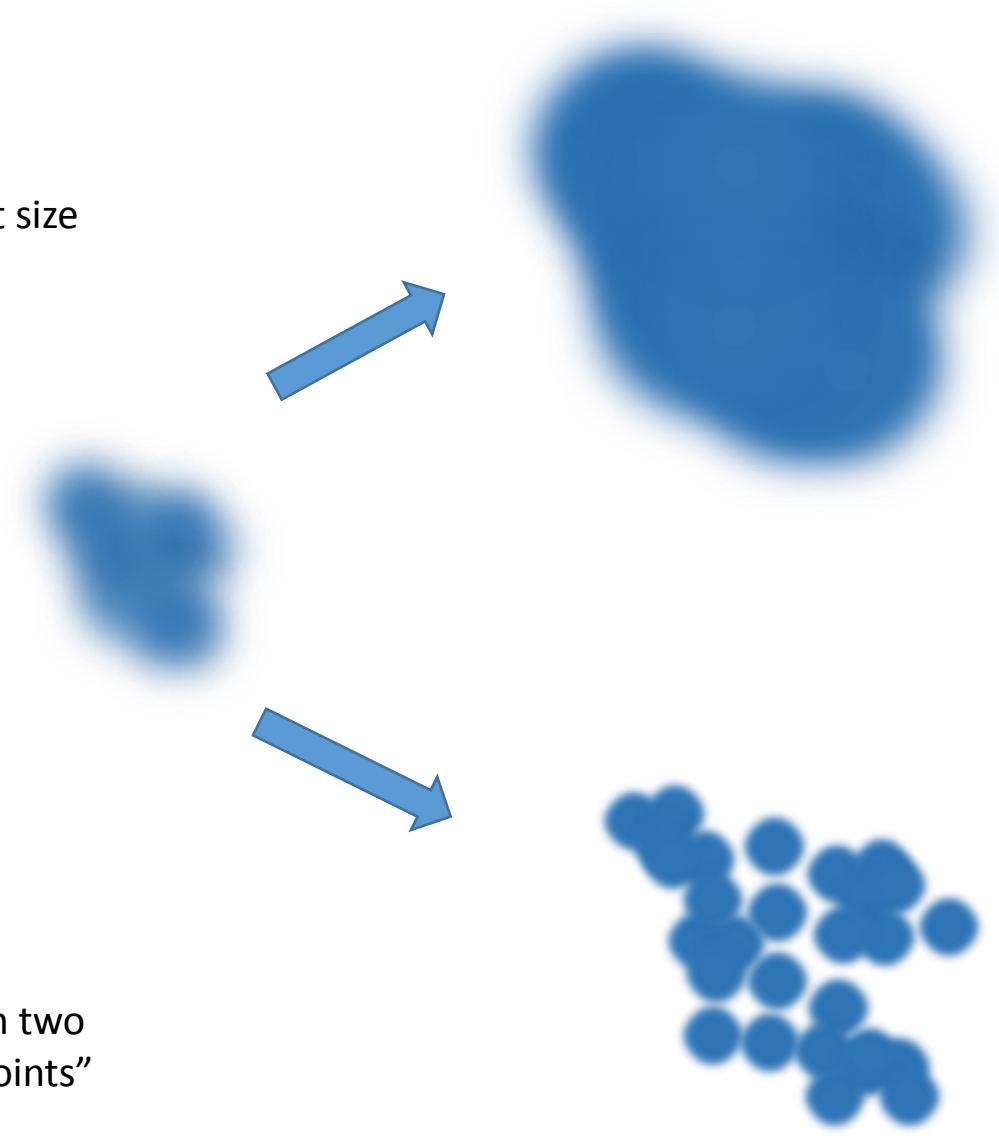
**TO SEE THINGS BIGGER**



## Resolution

“The smallest distance at which two points can be identify as two points”

**TO SEE MORE DETAILS**

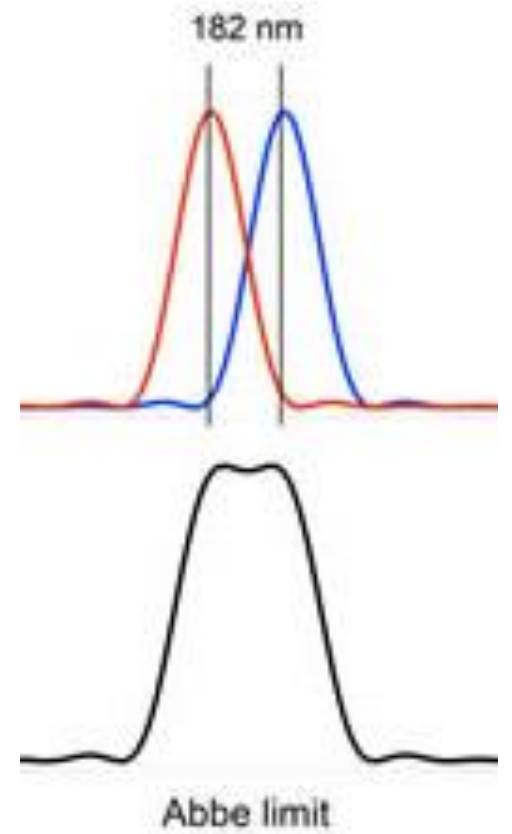
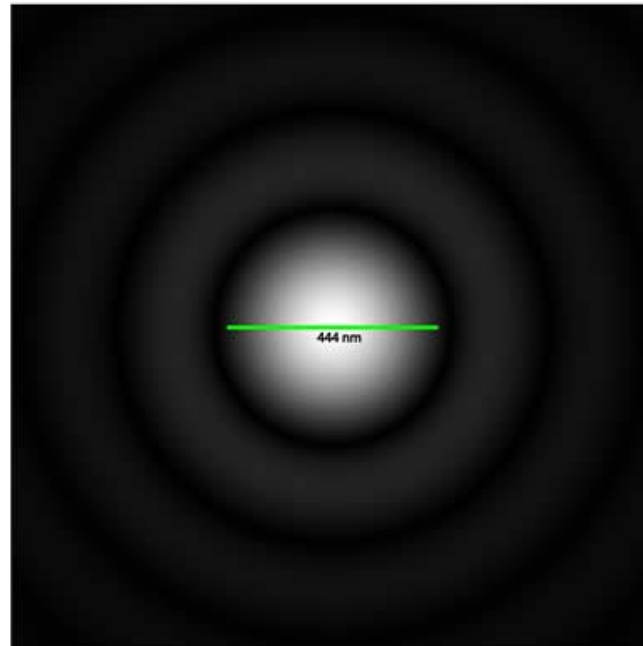
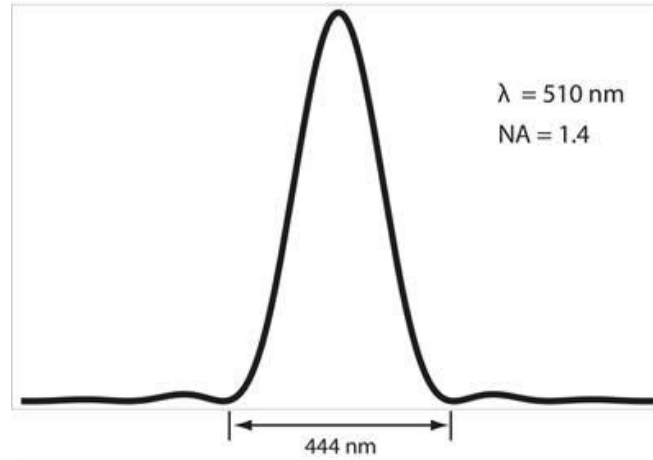
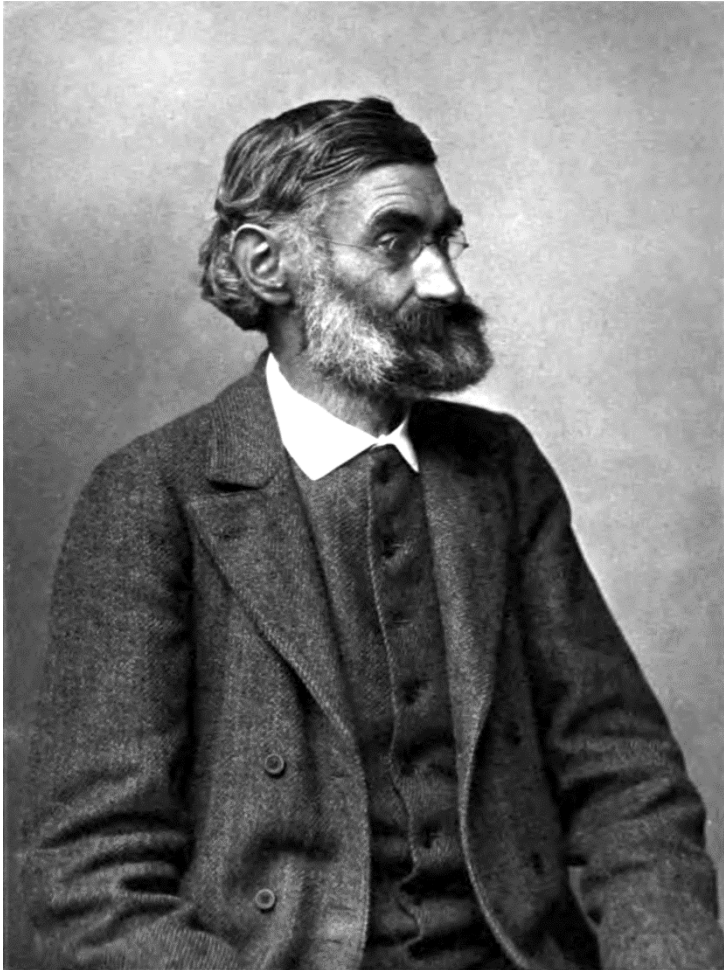


resolution



# Ernst Abbe Resolution barrier

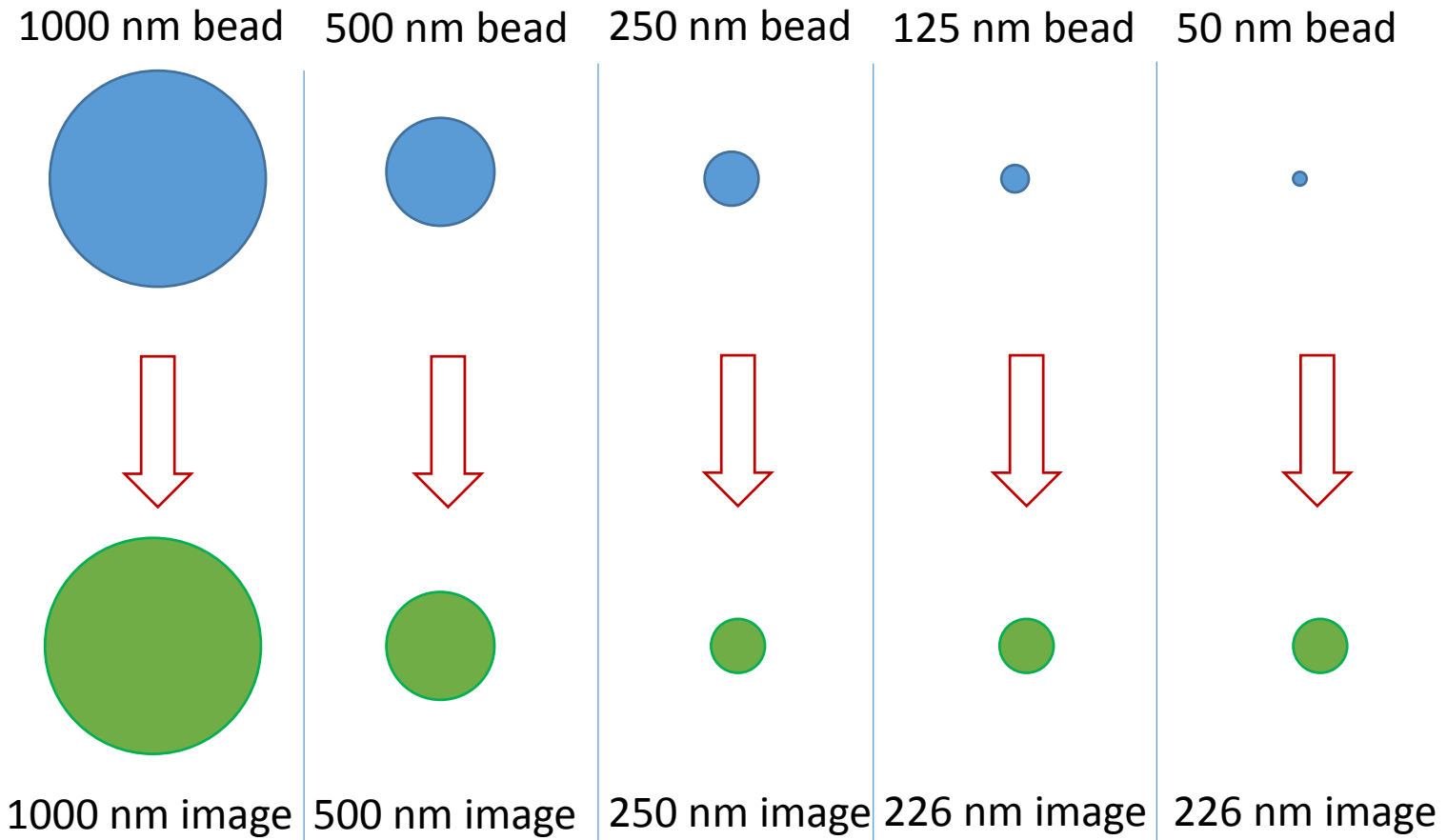
$$d = \frac{\lambda}{2NA}$$



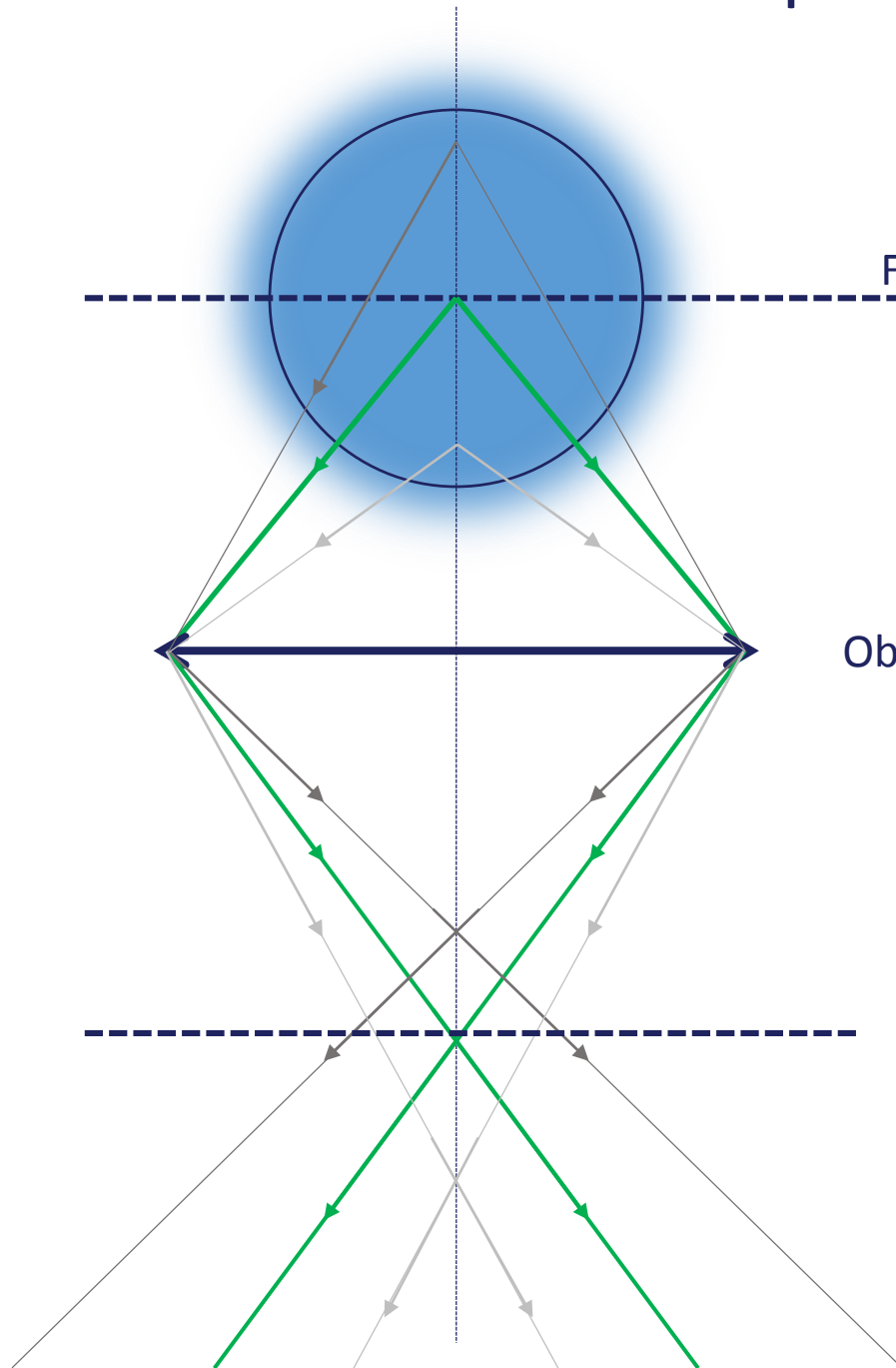
$$d = \frac{\lambda}{2NA}$$



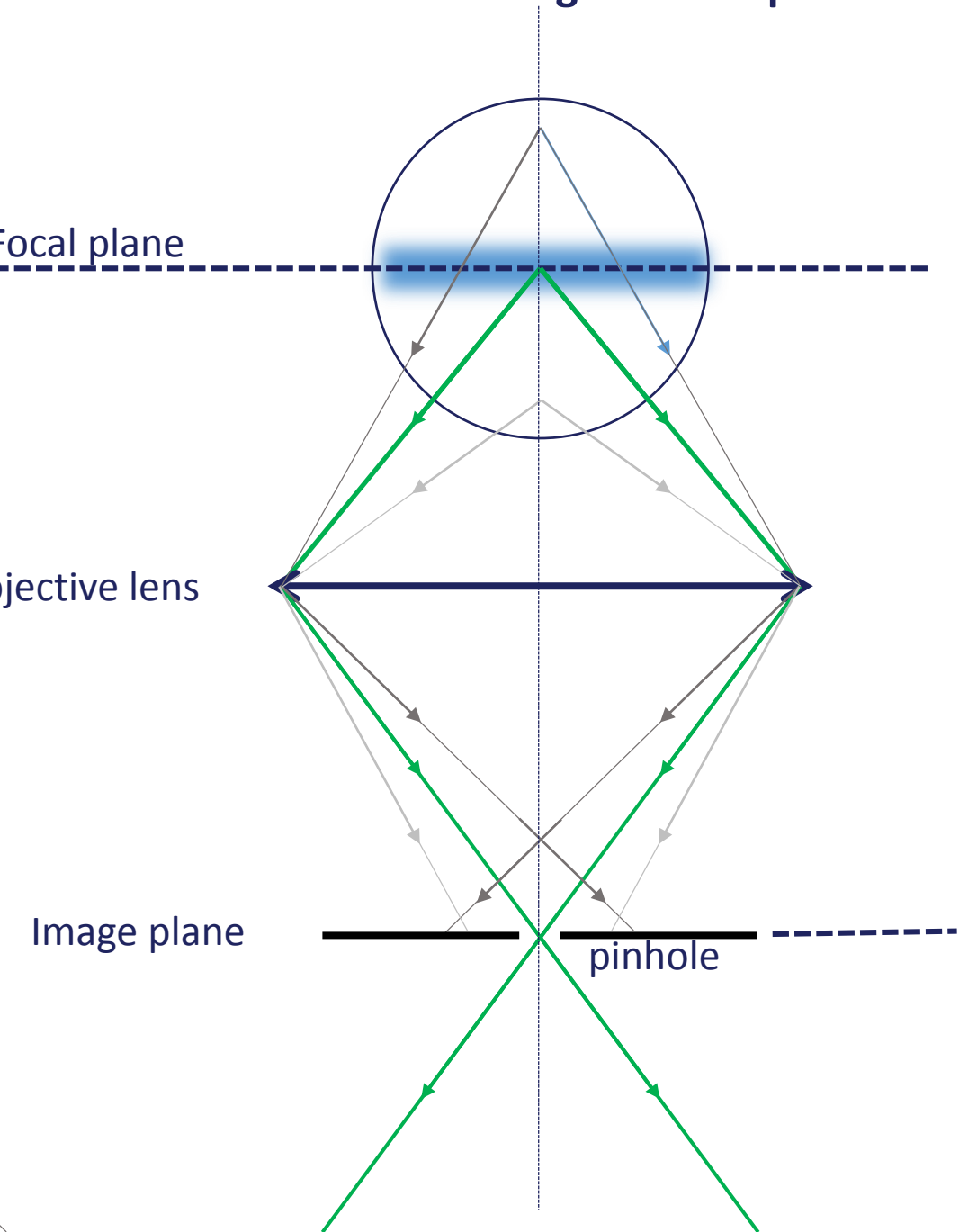
e.g., for  $\lambda = 520$  and  $NA = 1.4$ , the resolution is 226.6 nm:



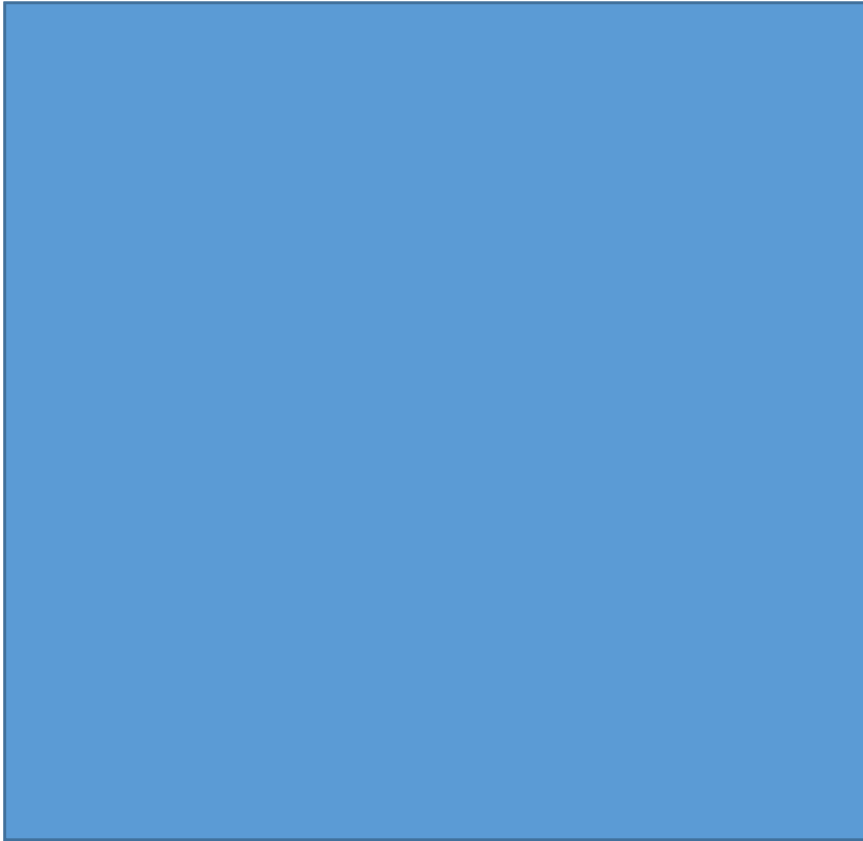
# Wide Field Fluorescent microscope



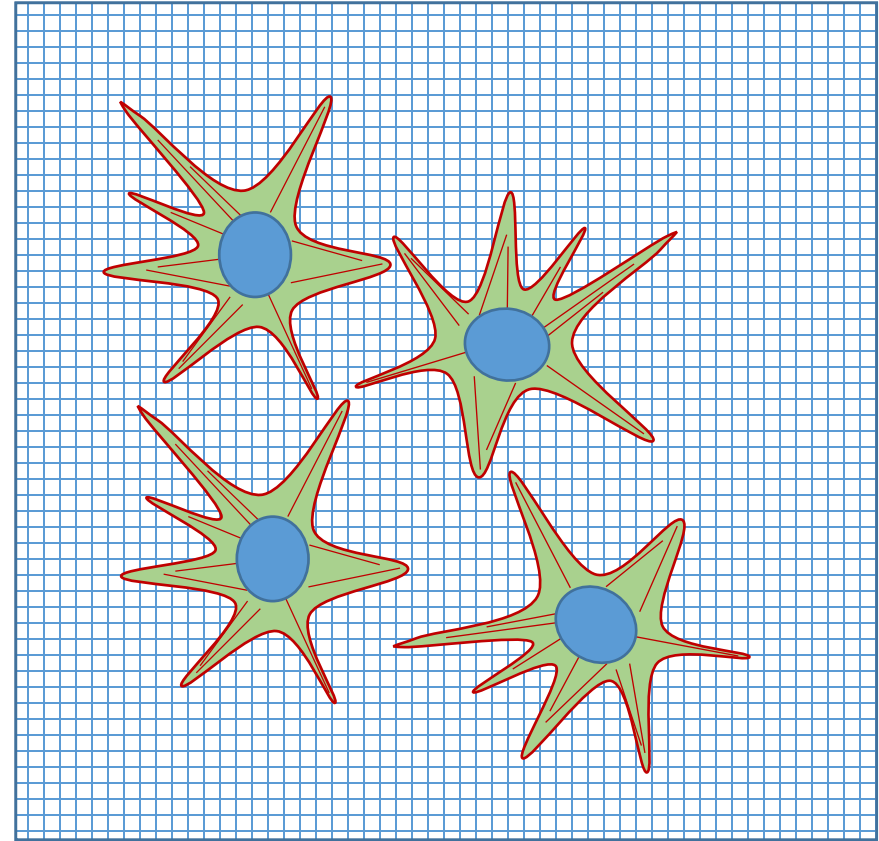
# Confocal scanning microscope



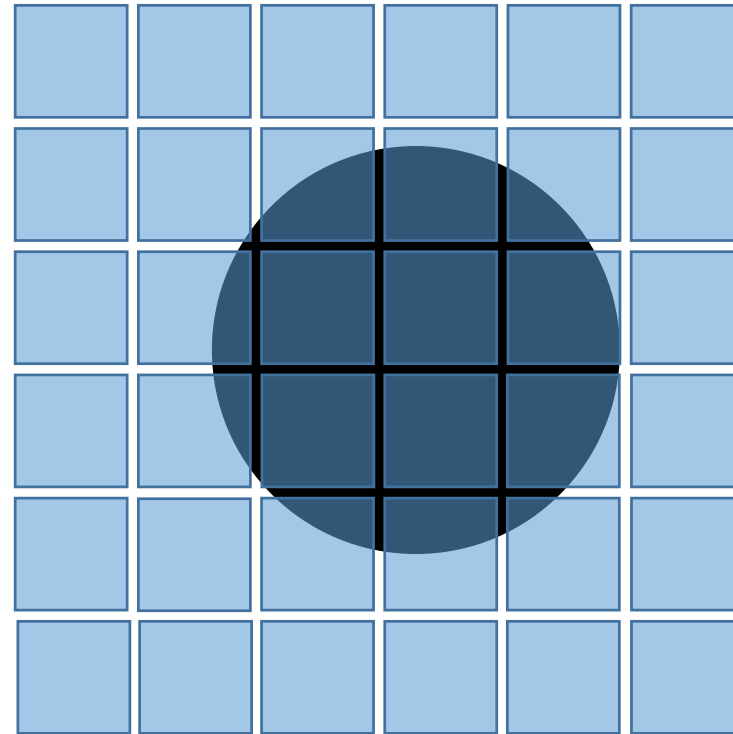
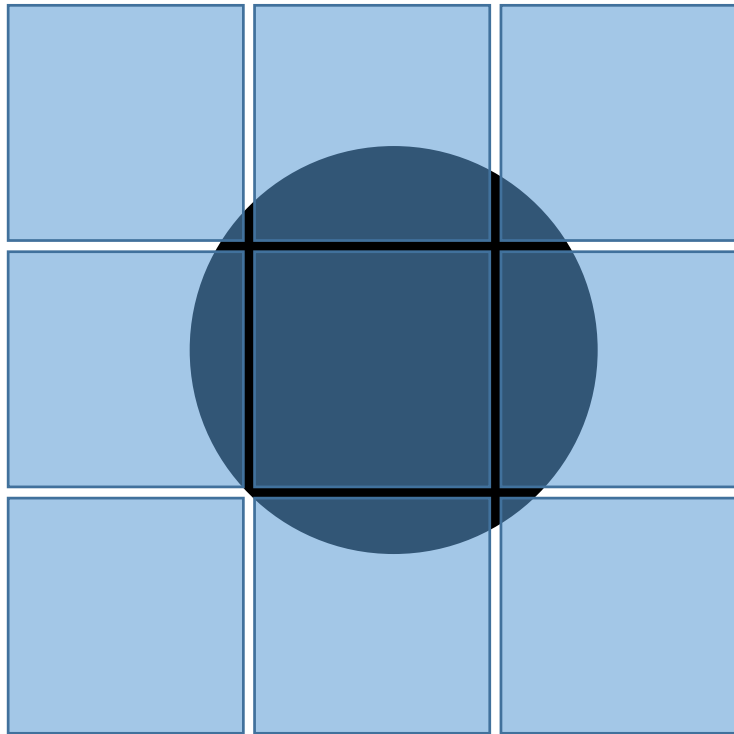
**Wide field microscope**



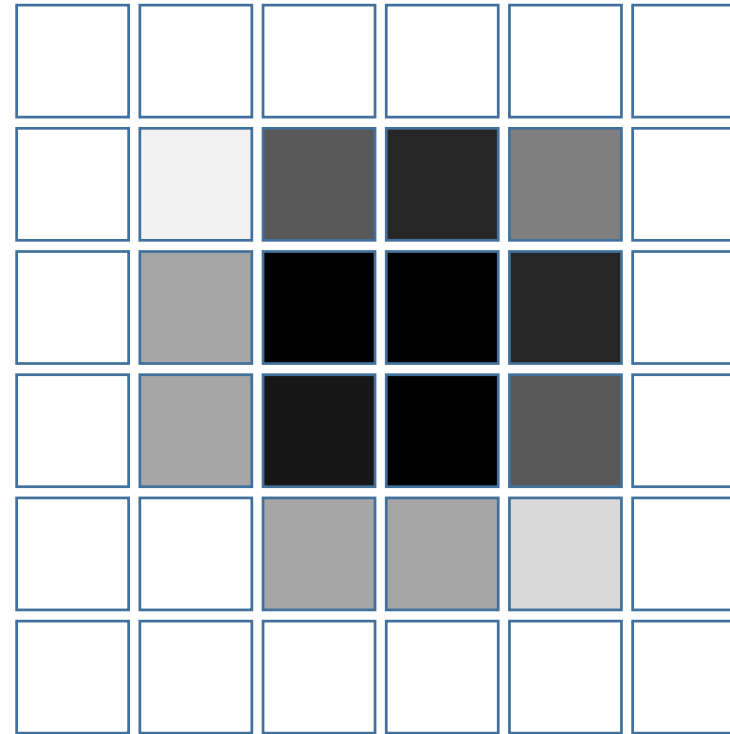
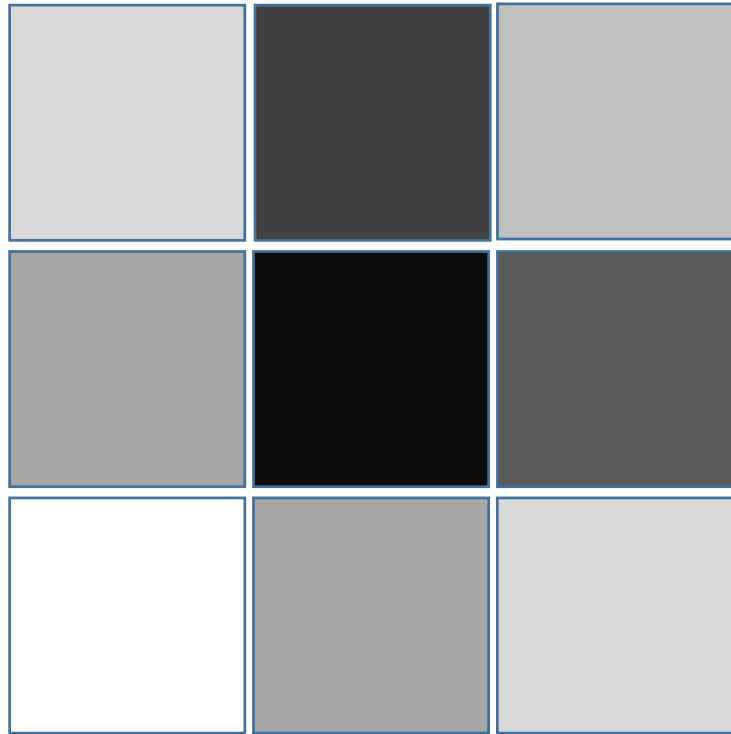
**Confocal scanning microscope**



# Confocal scanning microscope



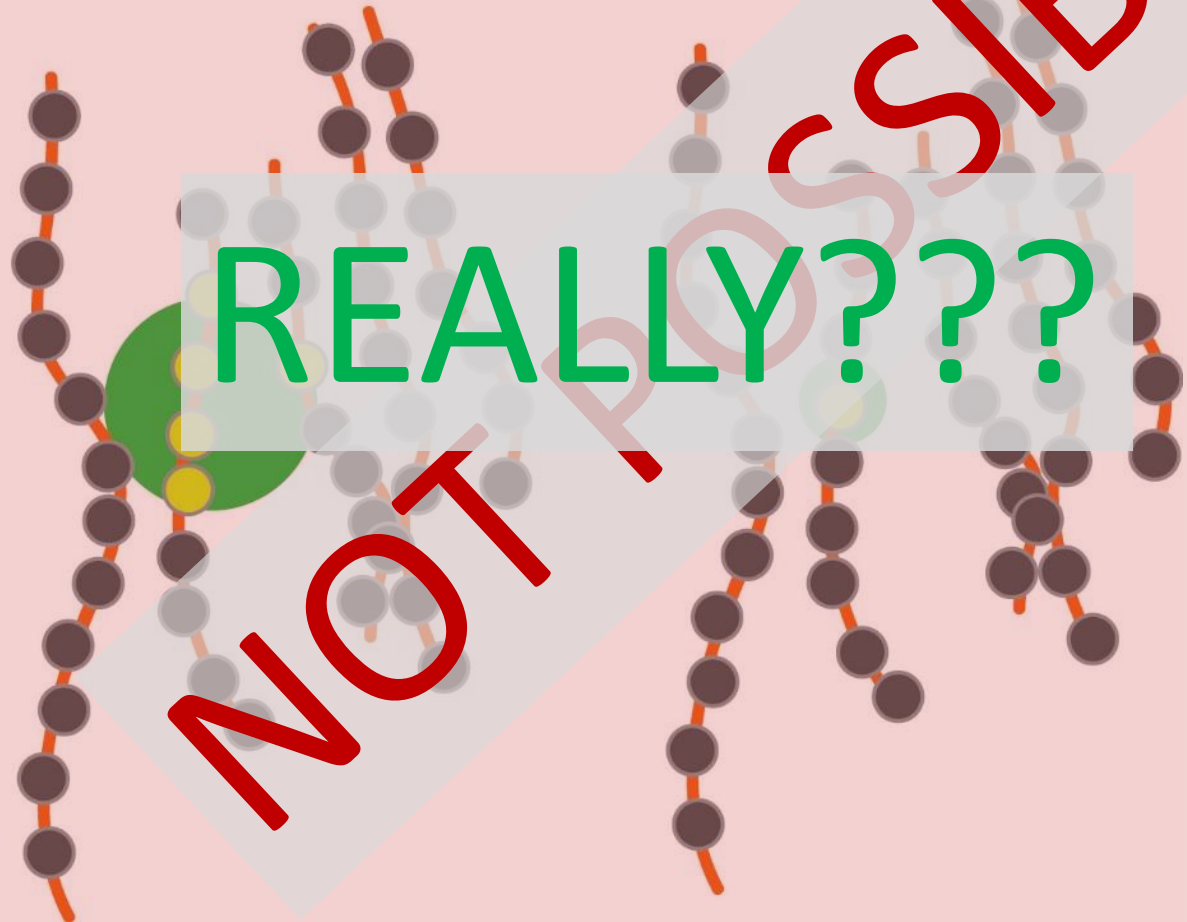
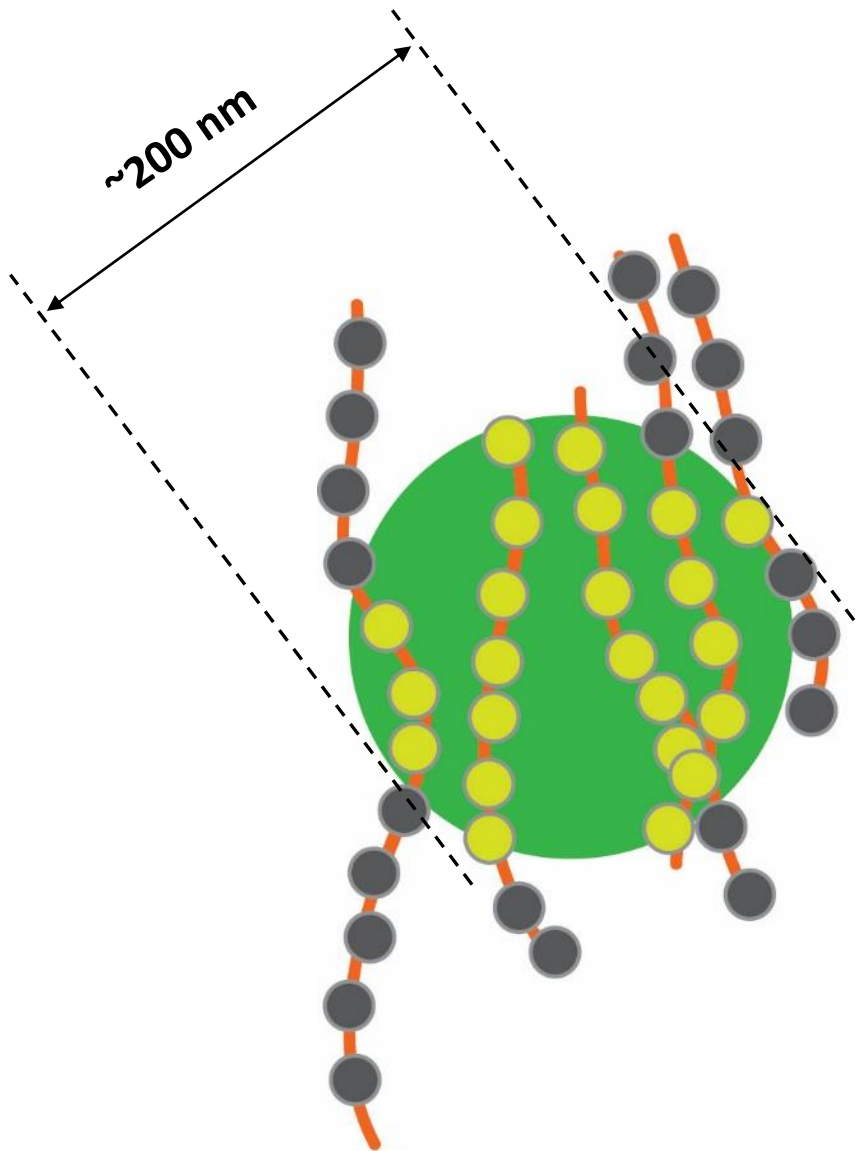
# Confocal scanning microscope



Ernst Abbe

Resolution barrier

$$d = \frac{\lambda}{2NA}$$



NOT POSSIBLE



**Stefan W. Hell**

**Born:** 23 December 1962, Arad,  
Romania

**Affiliation at the time of the  
award:** Max Planck Institute for  
Biophysical Chemistry, Göttingen,  
Germany, German Cancer Research  
Center, Heidelberg, Germany

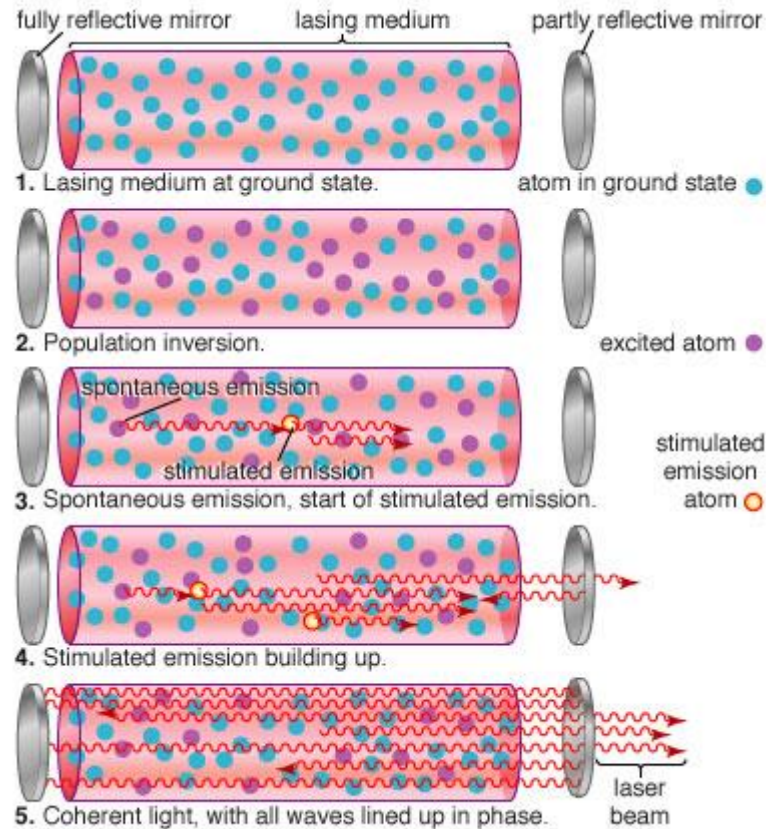
**Prize motivation:** "for the  
development of super-resolved  
fluorescence microscopy"

**Field:** physical chemistry

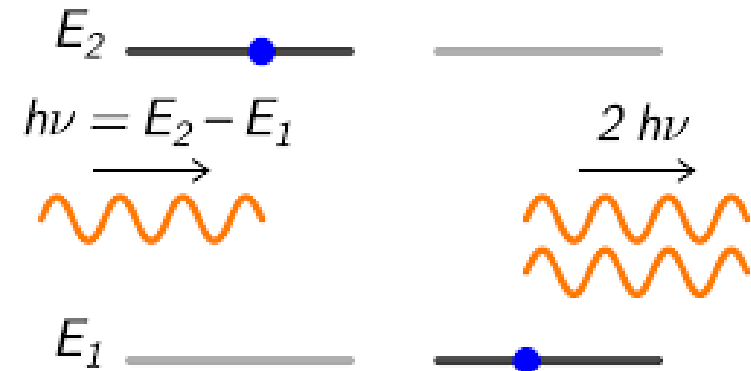
**Prize share:** 1/3

*[https://www.nobelprize.org/nobel\\_prizes/chemistry/laureates/2014/hell-facts.html](https://www.nobelprize.org/nobel_prizes/chemistry/laureates/2014/hell-facts.html)*

# Principle of the LASER – stimulated emission



LASER = light amplification by stimulated emission of radiation.

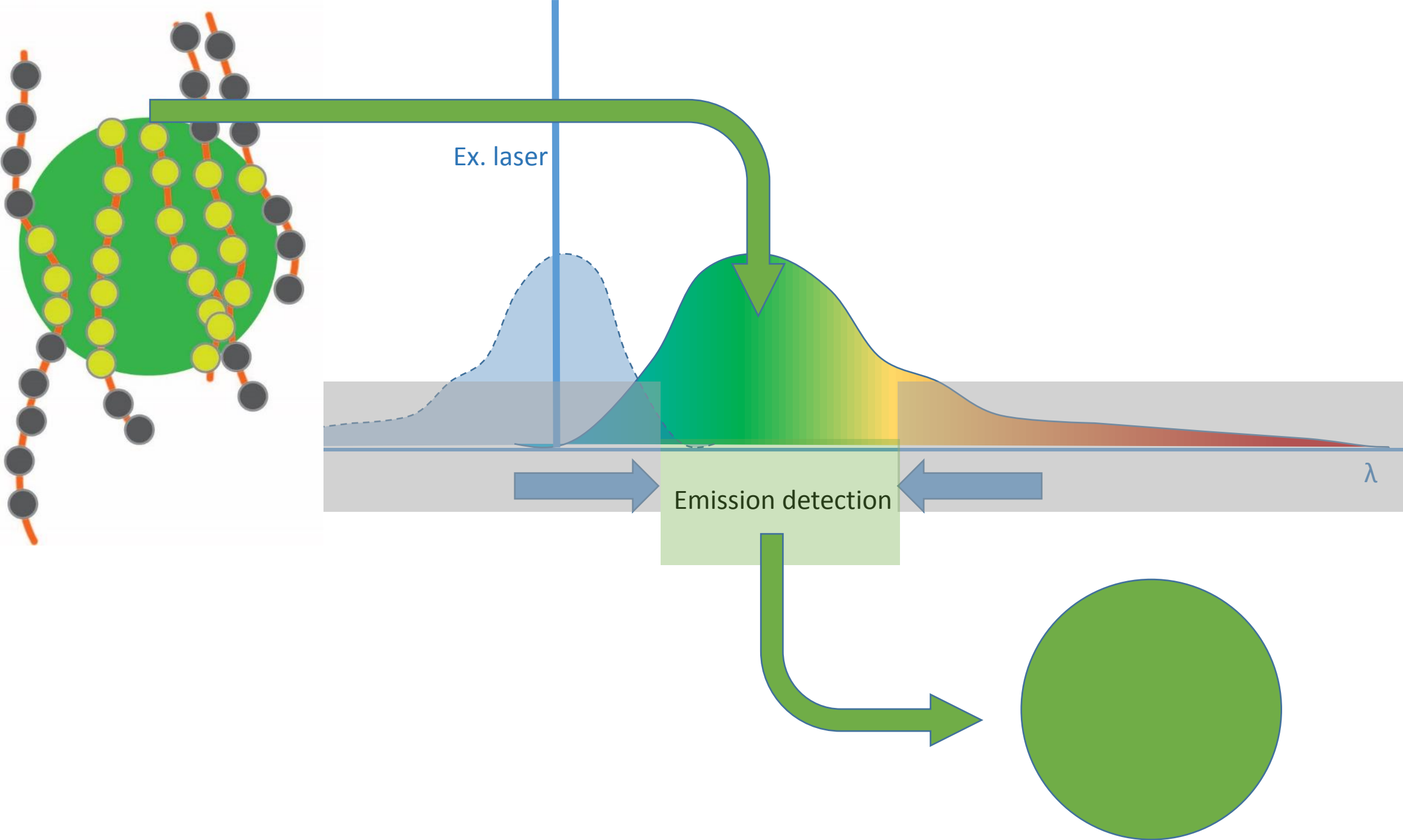


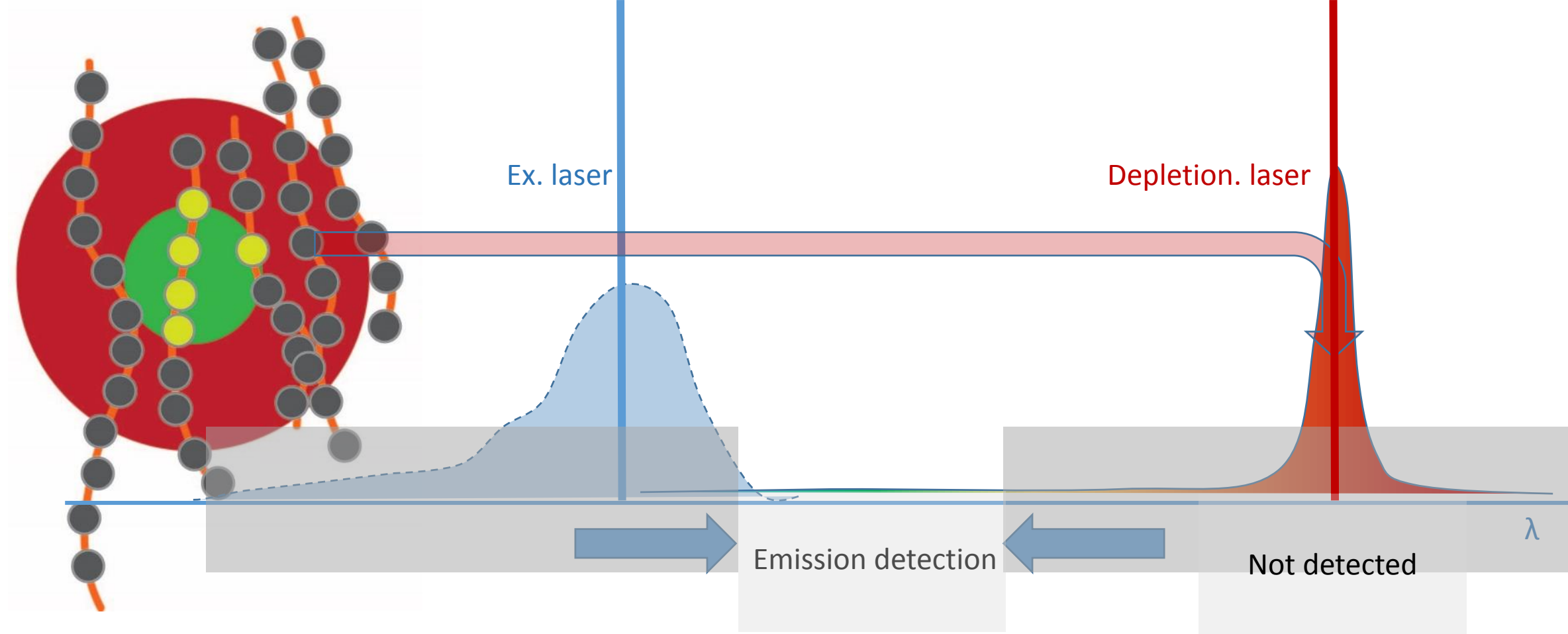
© 2006 Encyclopædia Britannica, Inc.

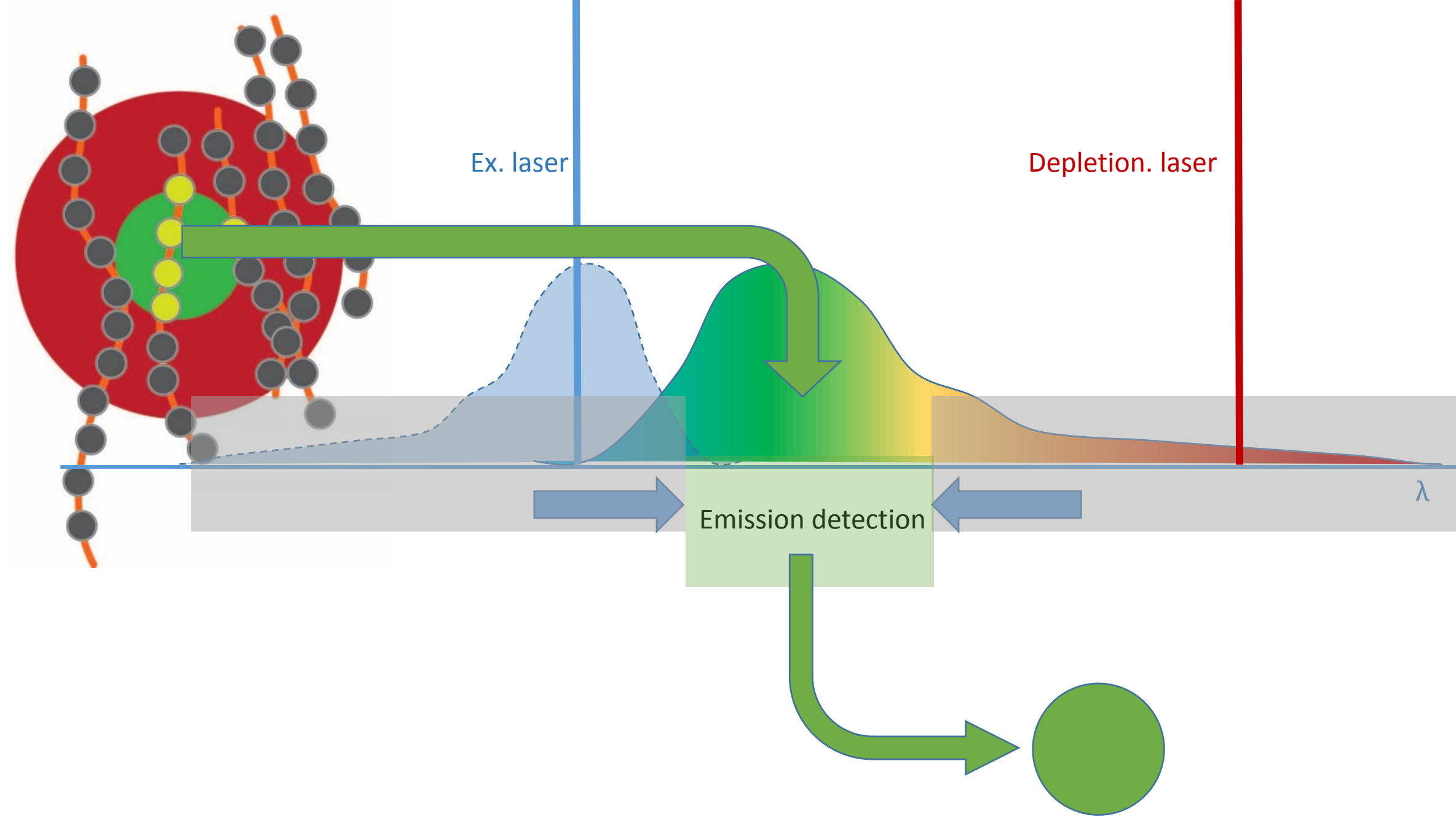
<https://www.britannica.com/technology/stimulated-emission>

<https://web2.ph.utexas.edu/~coker2/index.files/xrayslasers.htm>







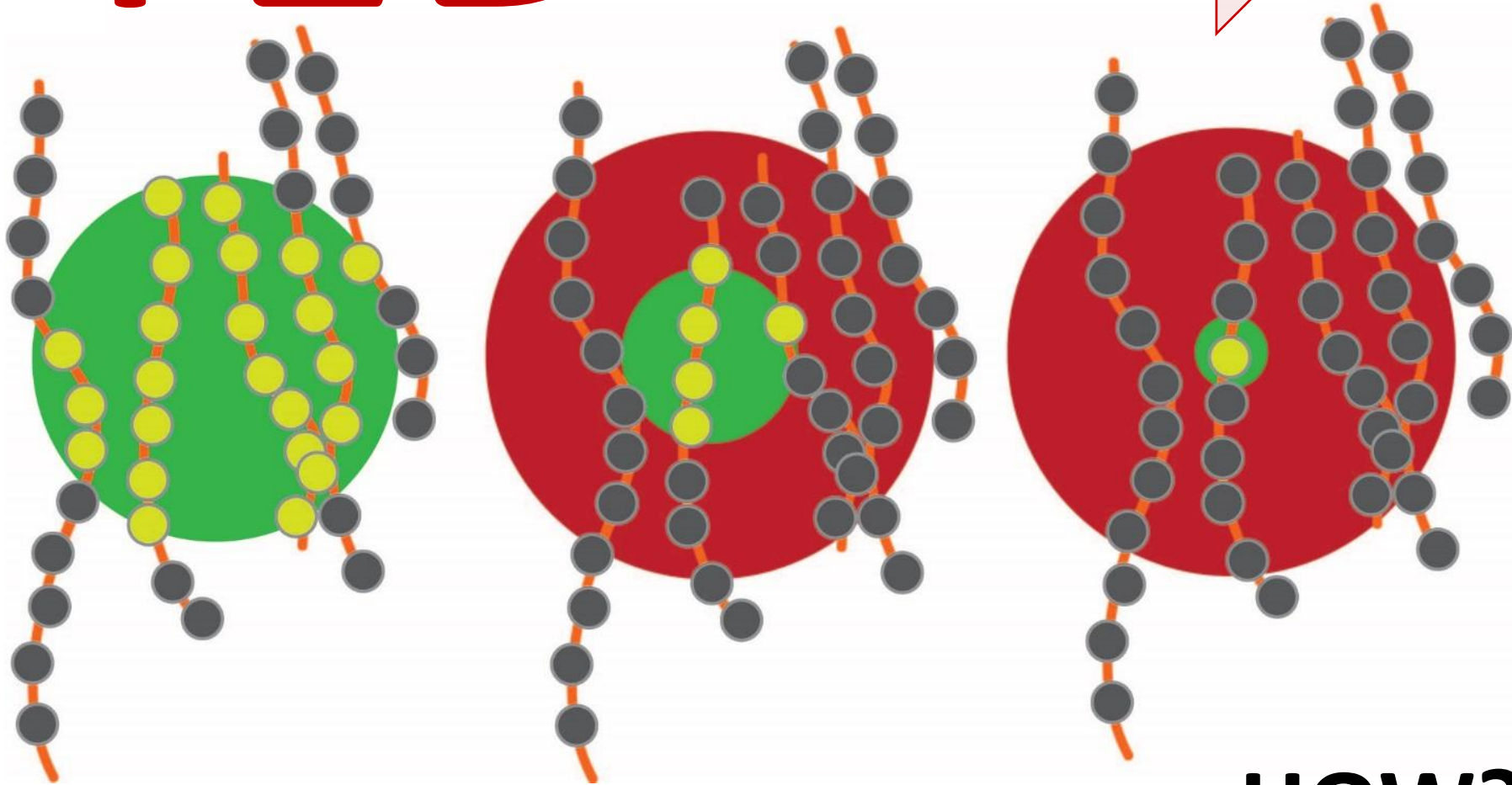


# STED

$$d = \frac{\lambda}{2NA}$$

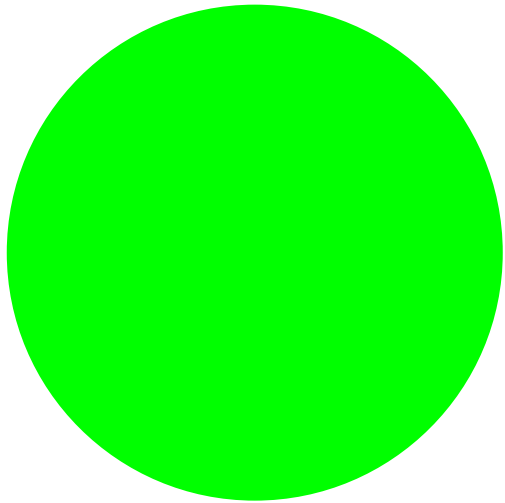
$$d_{STED} = \frac{\lambda}{2NA\sqrt{1 + \xi}}$$

break through the resolution barrier

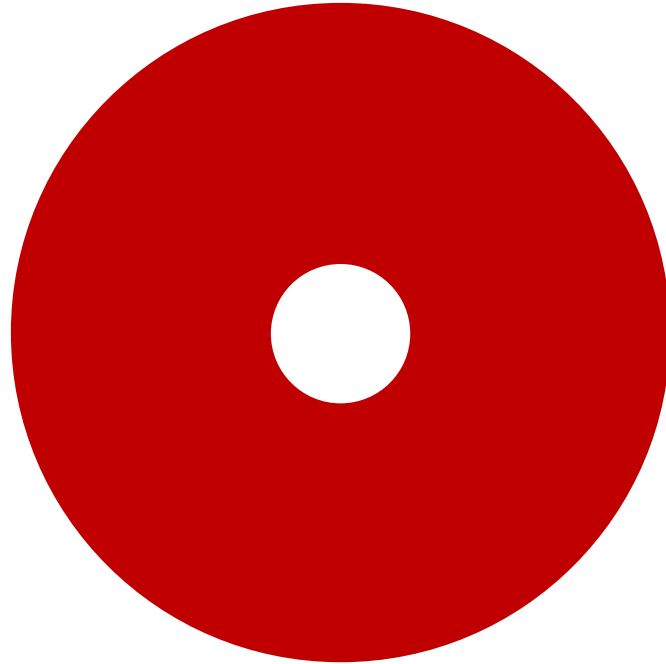


**HOW?**

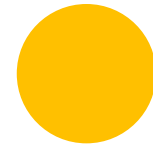
Excitation LASER spot

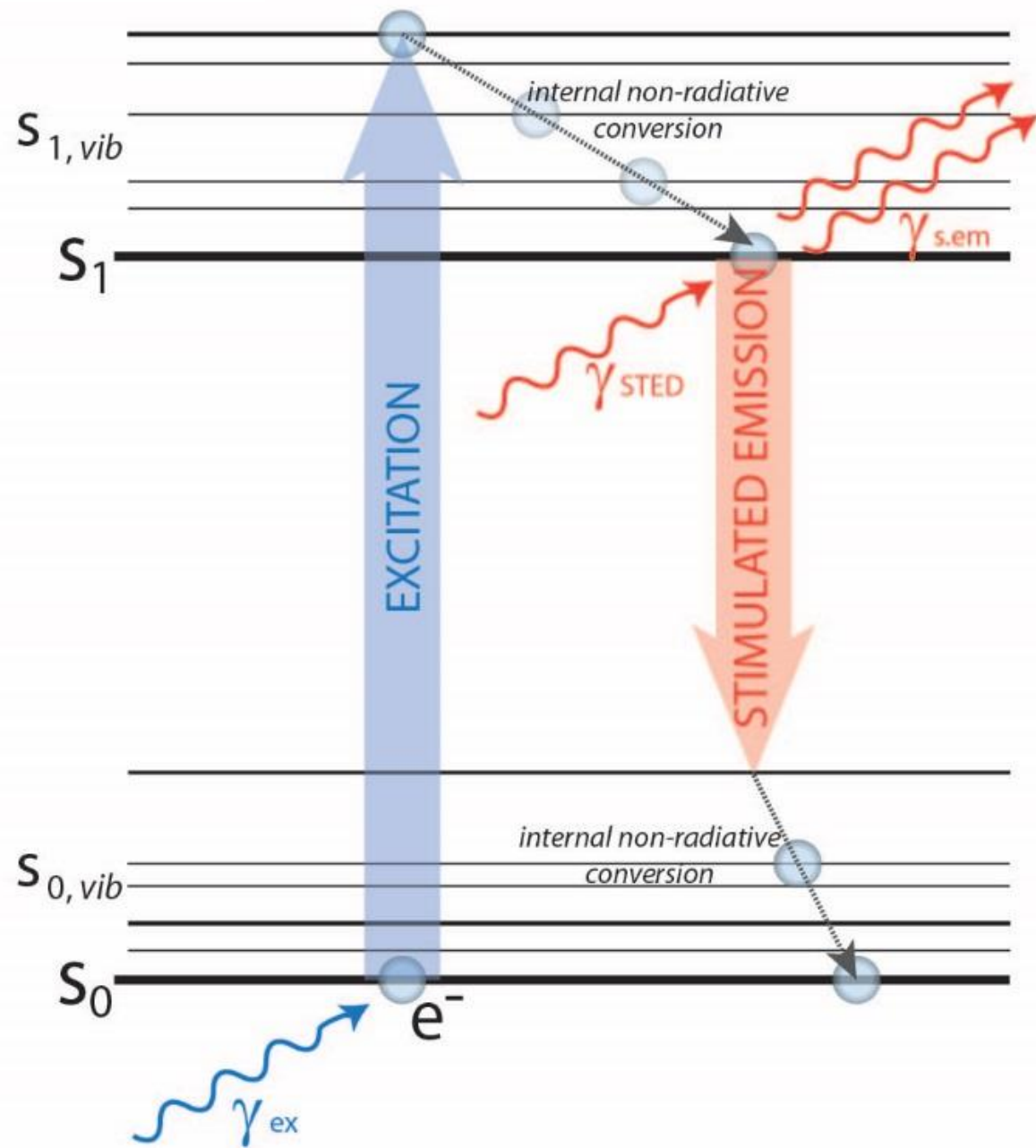
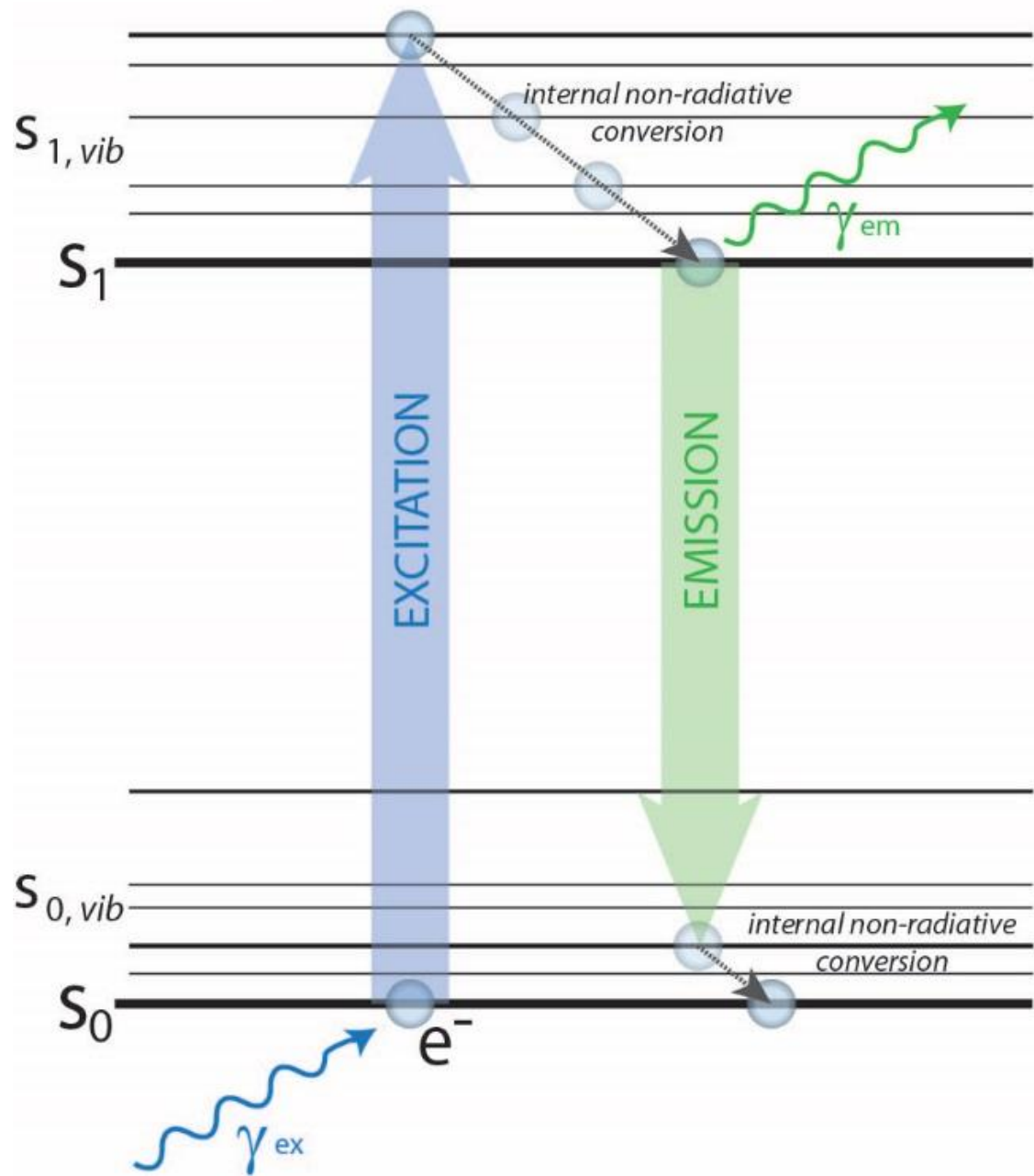


Depletion laser donut



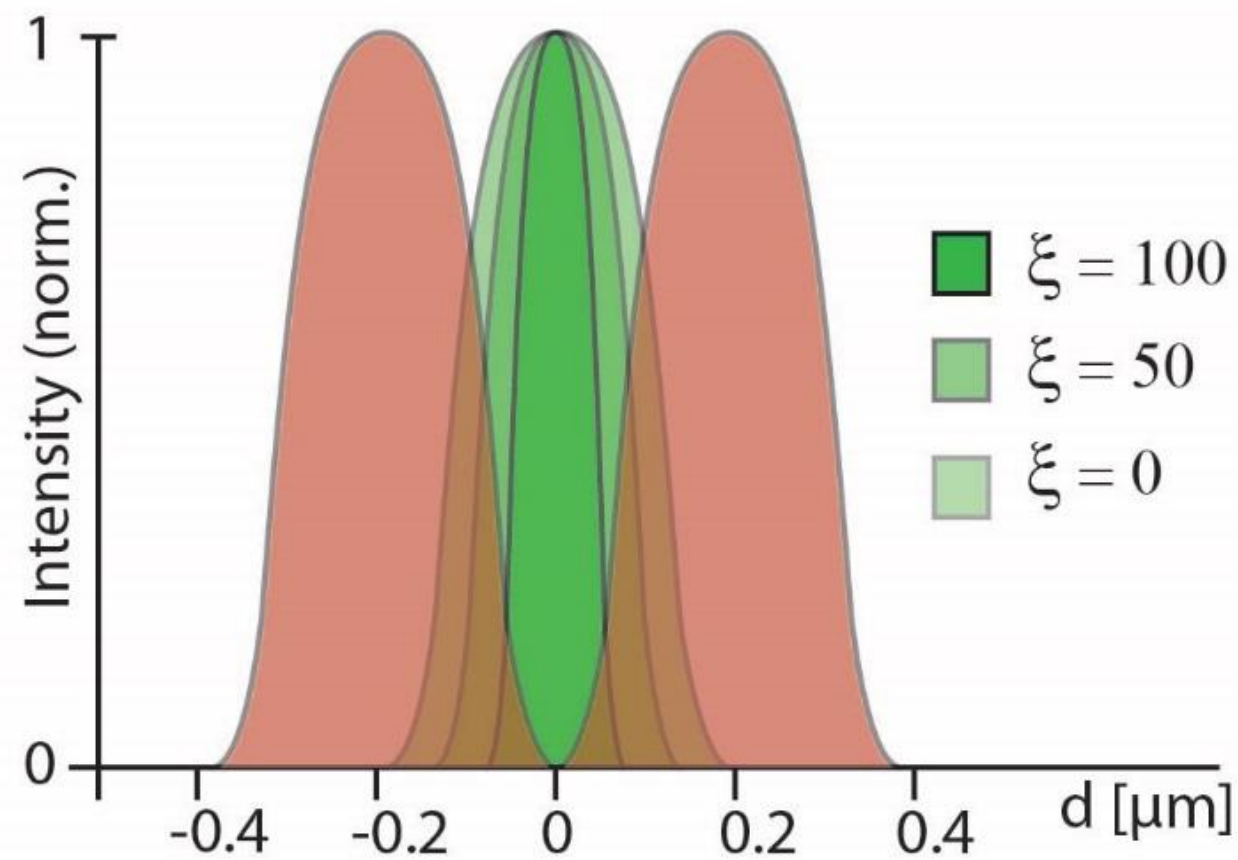
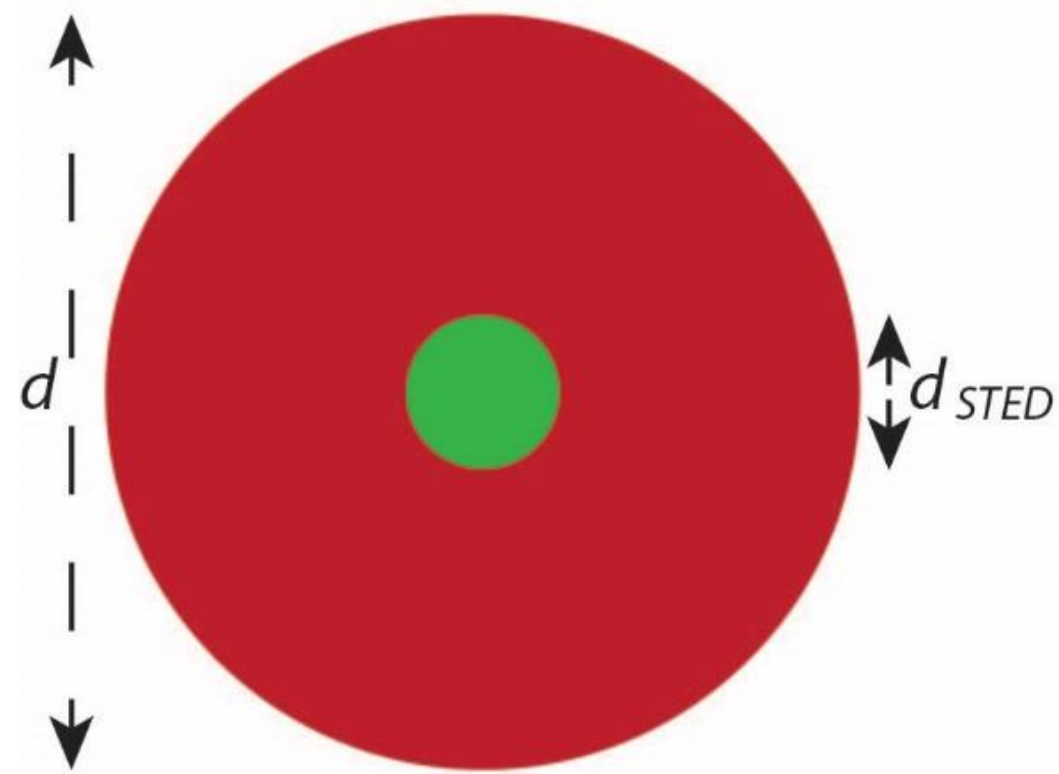
Acquired emission



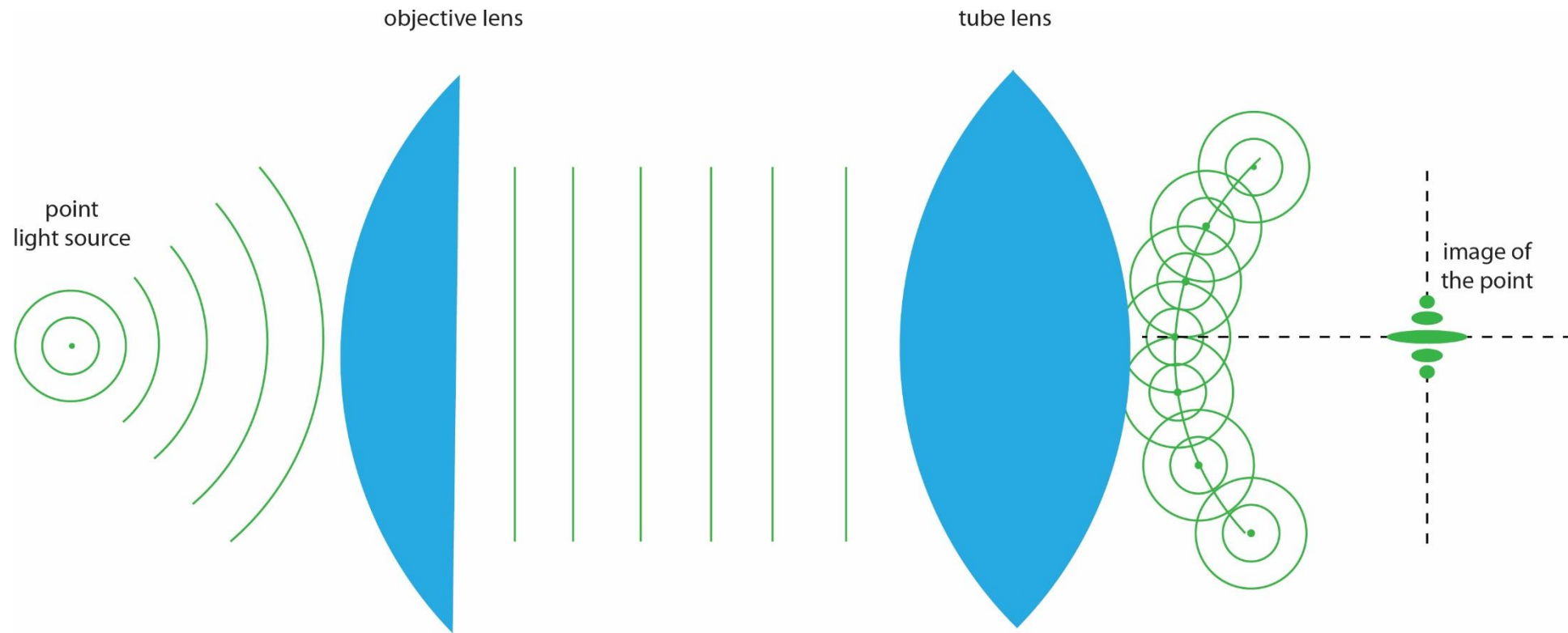


$$d_{STED} = \frac{\lambda}{2NA\sqrt{1+\xi}}$$

$$\xi = \frac{I_{STED}}{I_S}$$

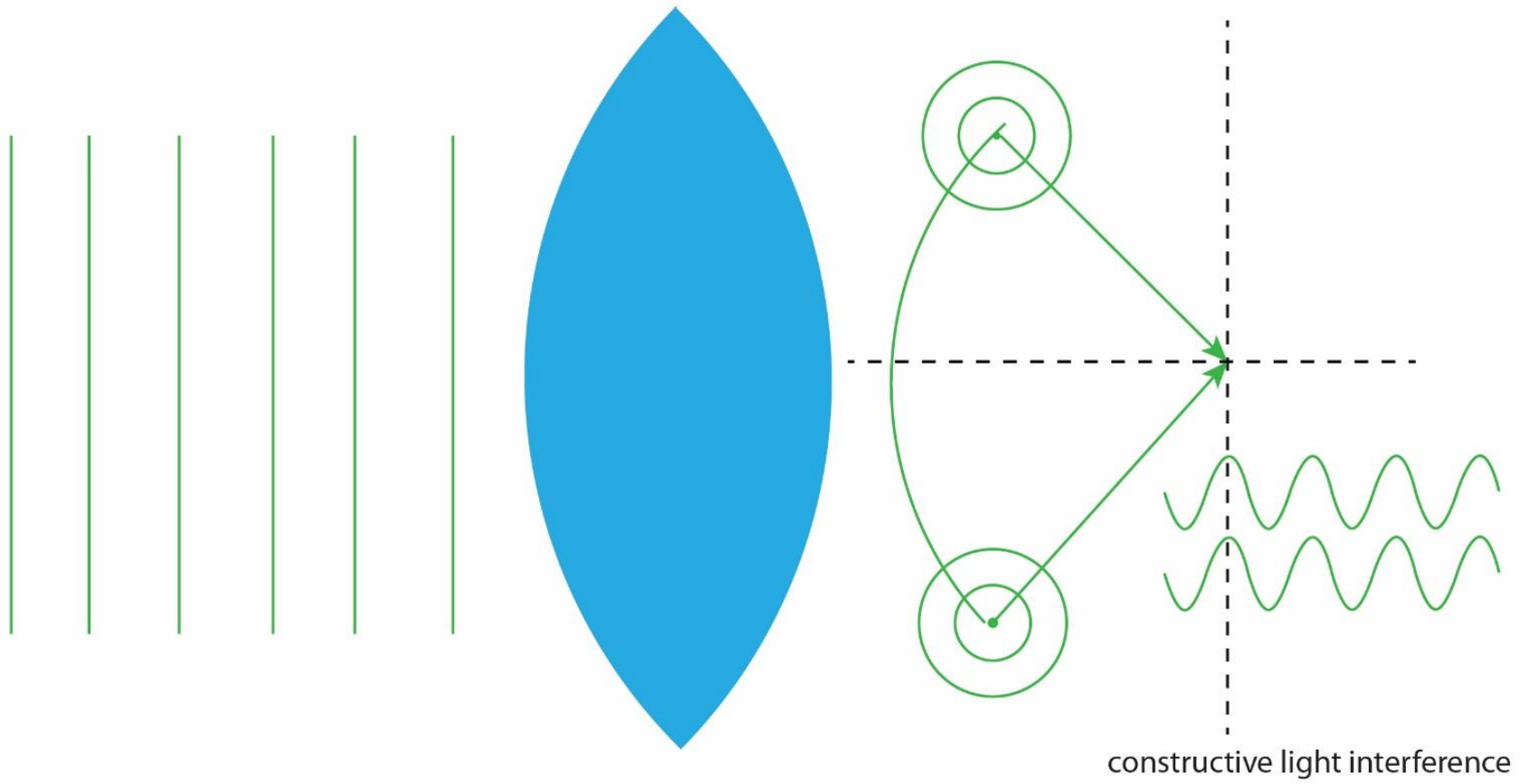


# How the donut is created?

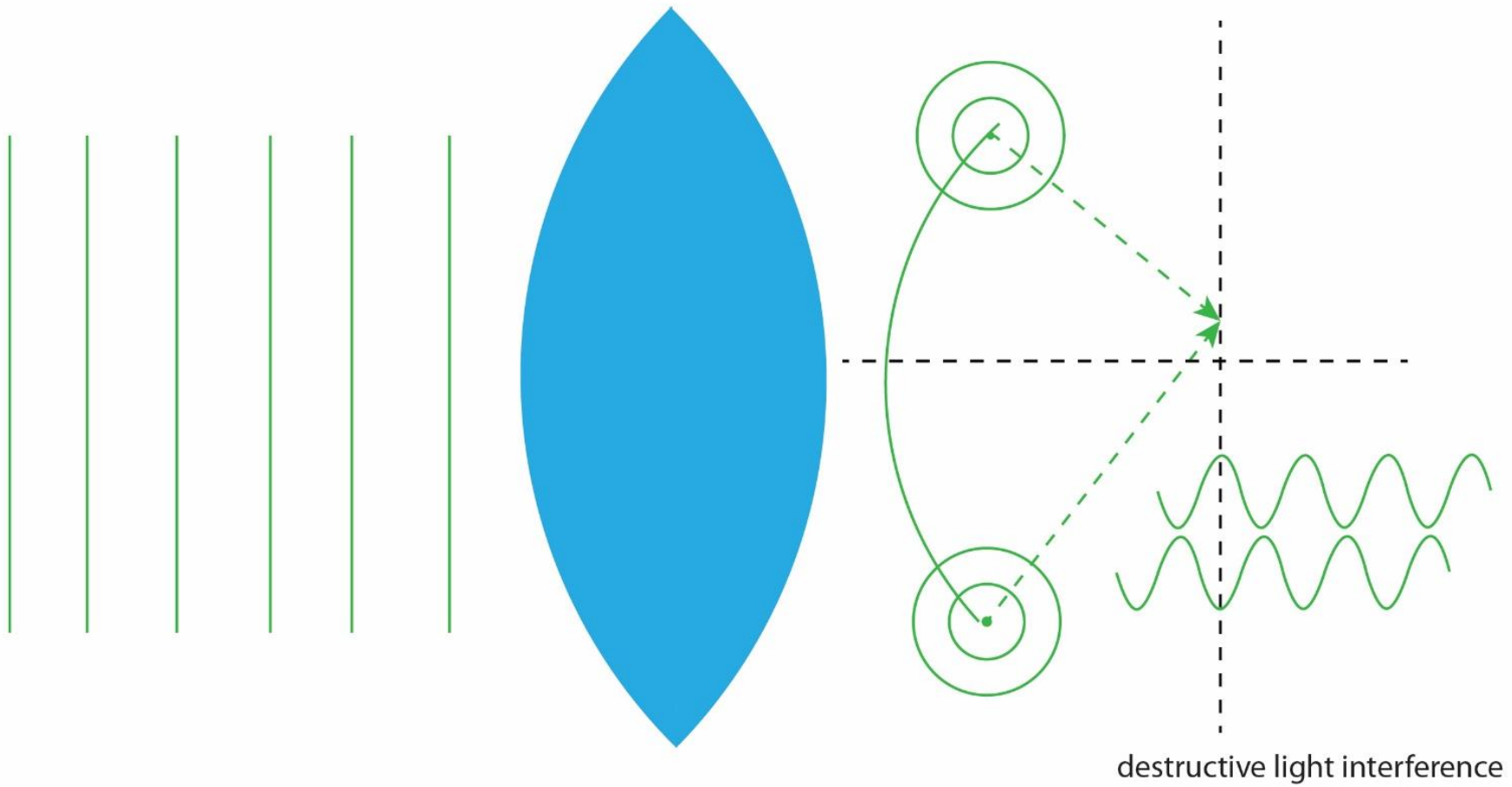




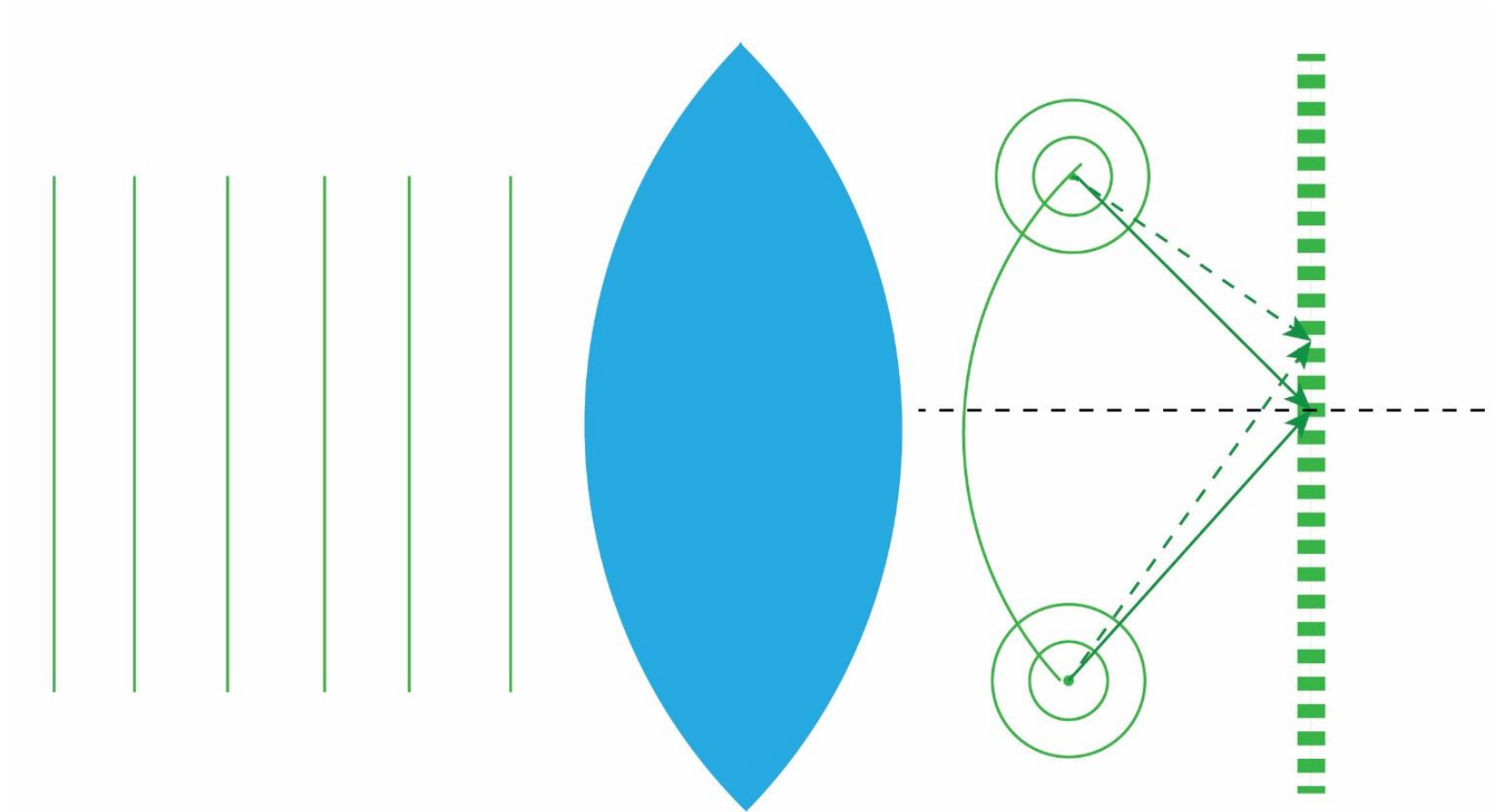
# How the donut is created?



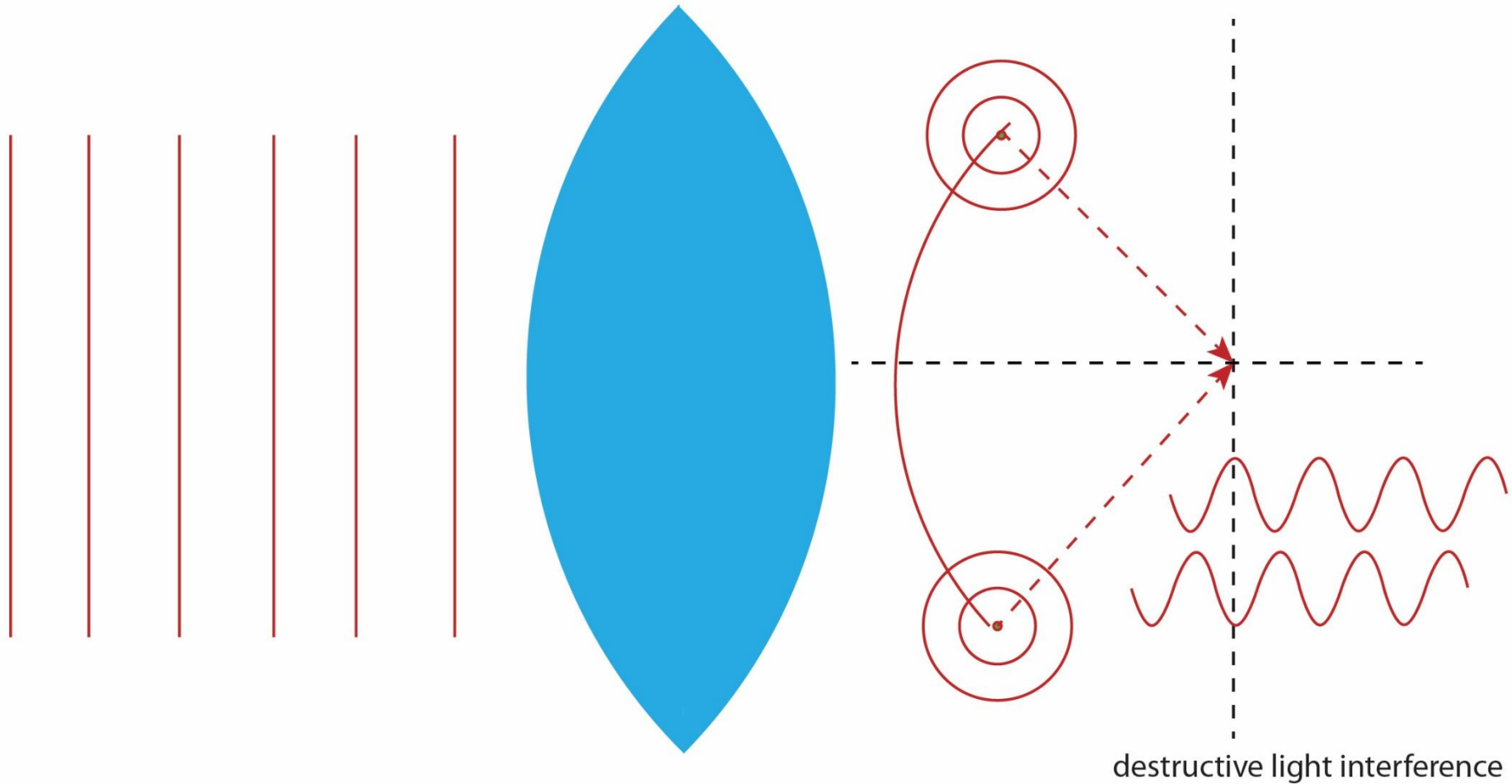
# How the donut is created?



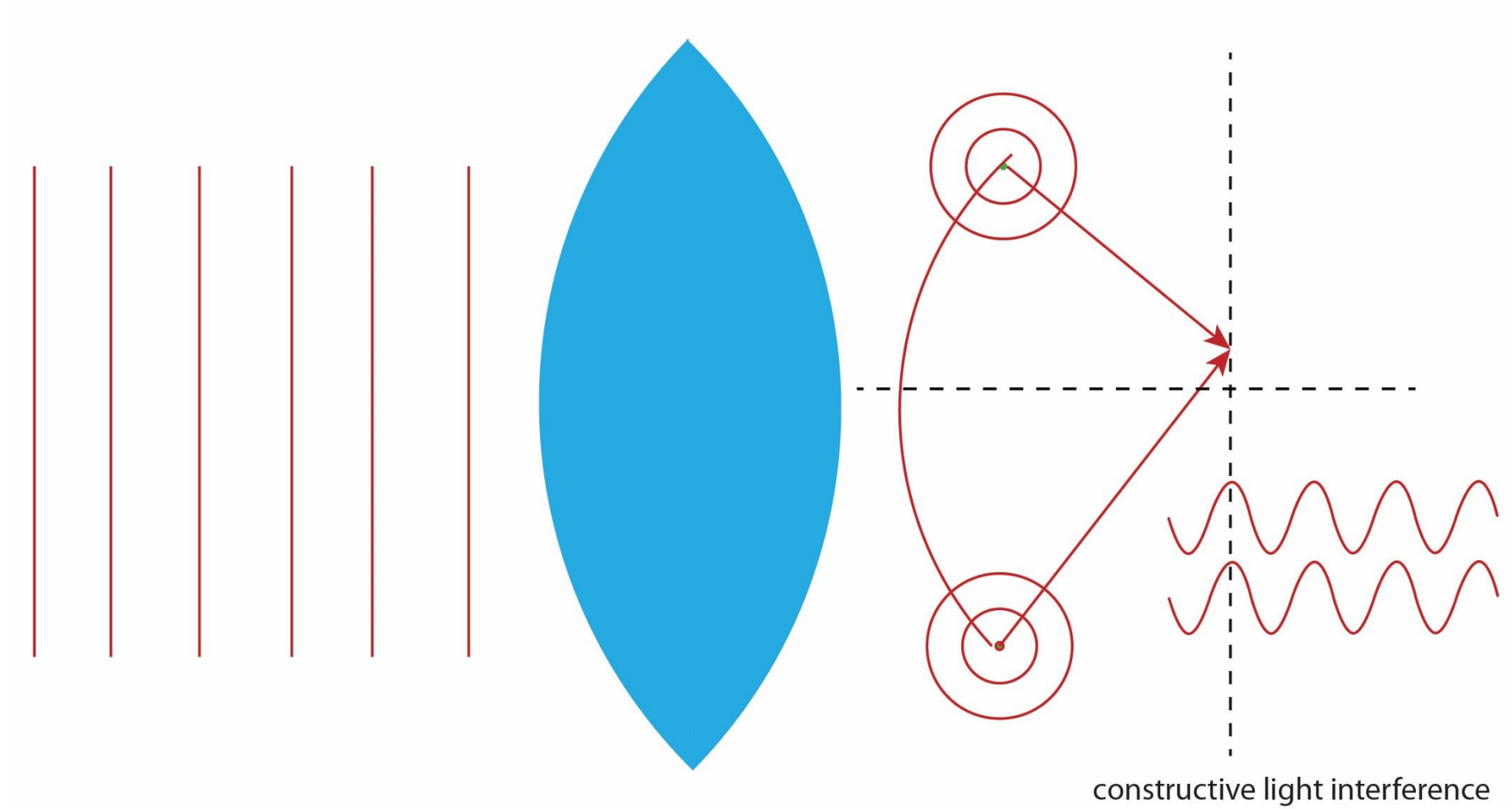
How the donut is created?



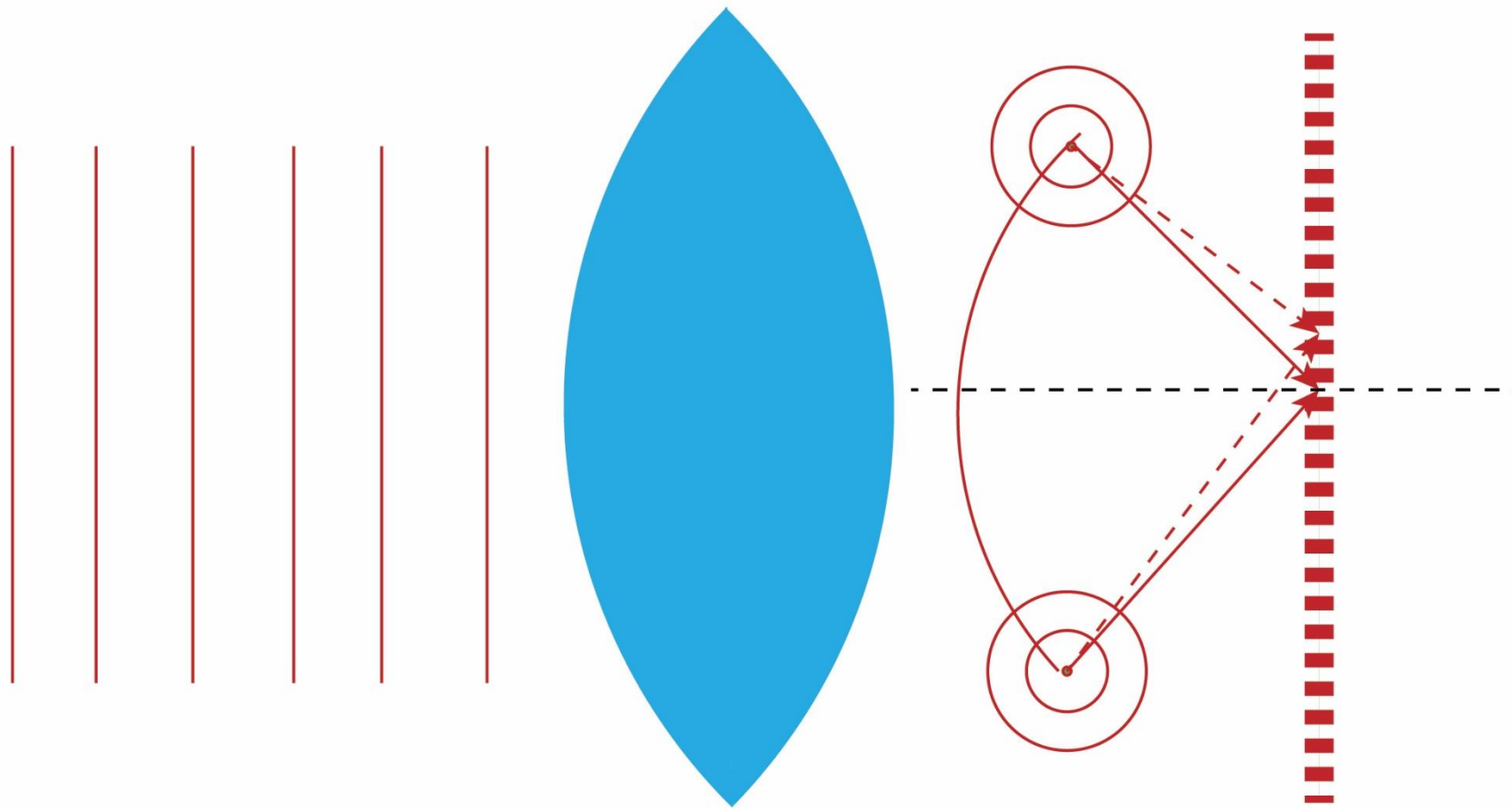
# How the donut is created?



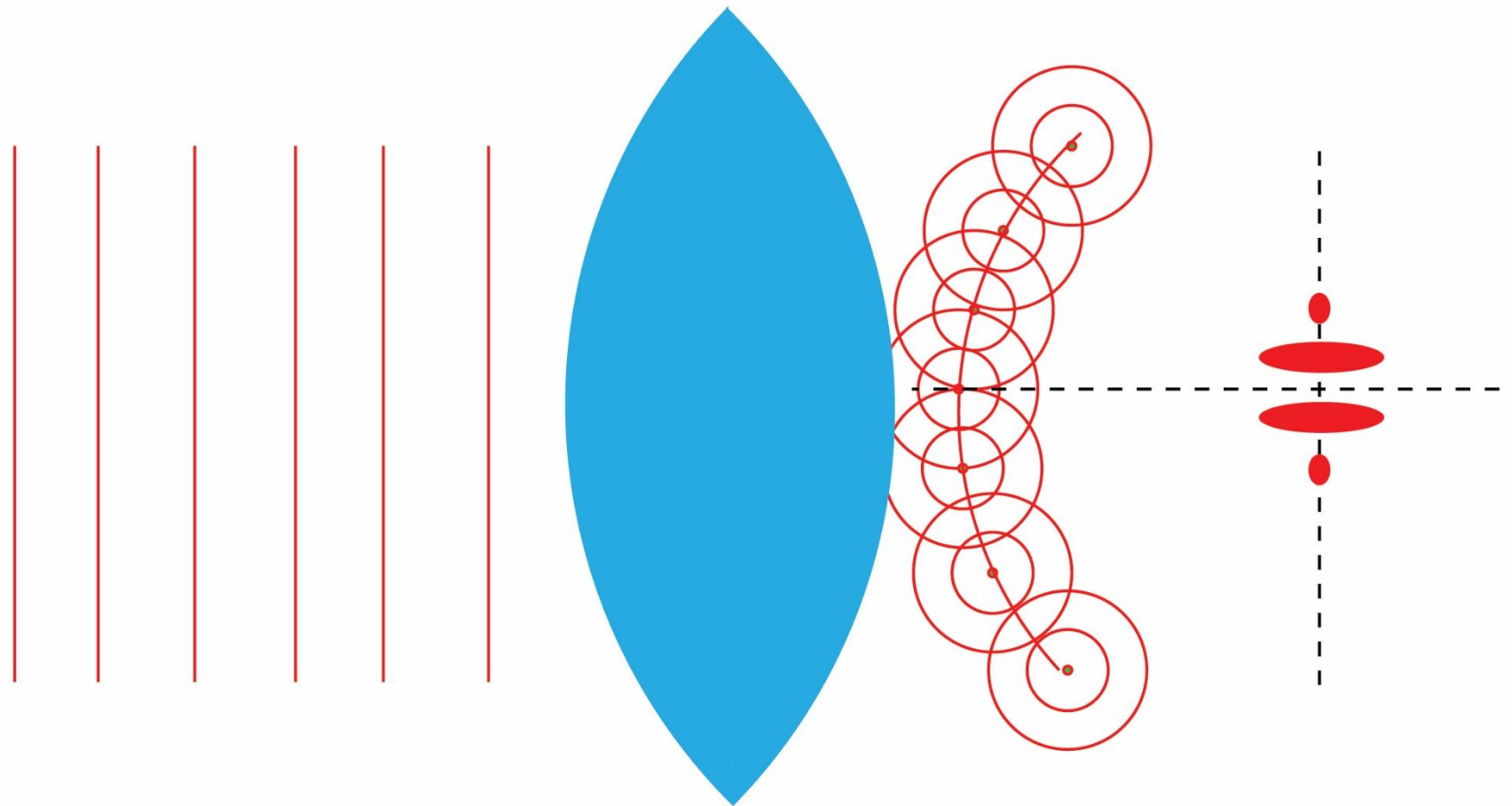
# How the donut is created?



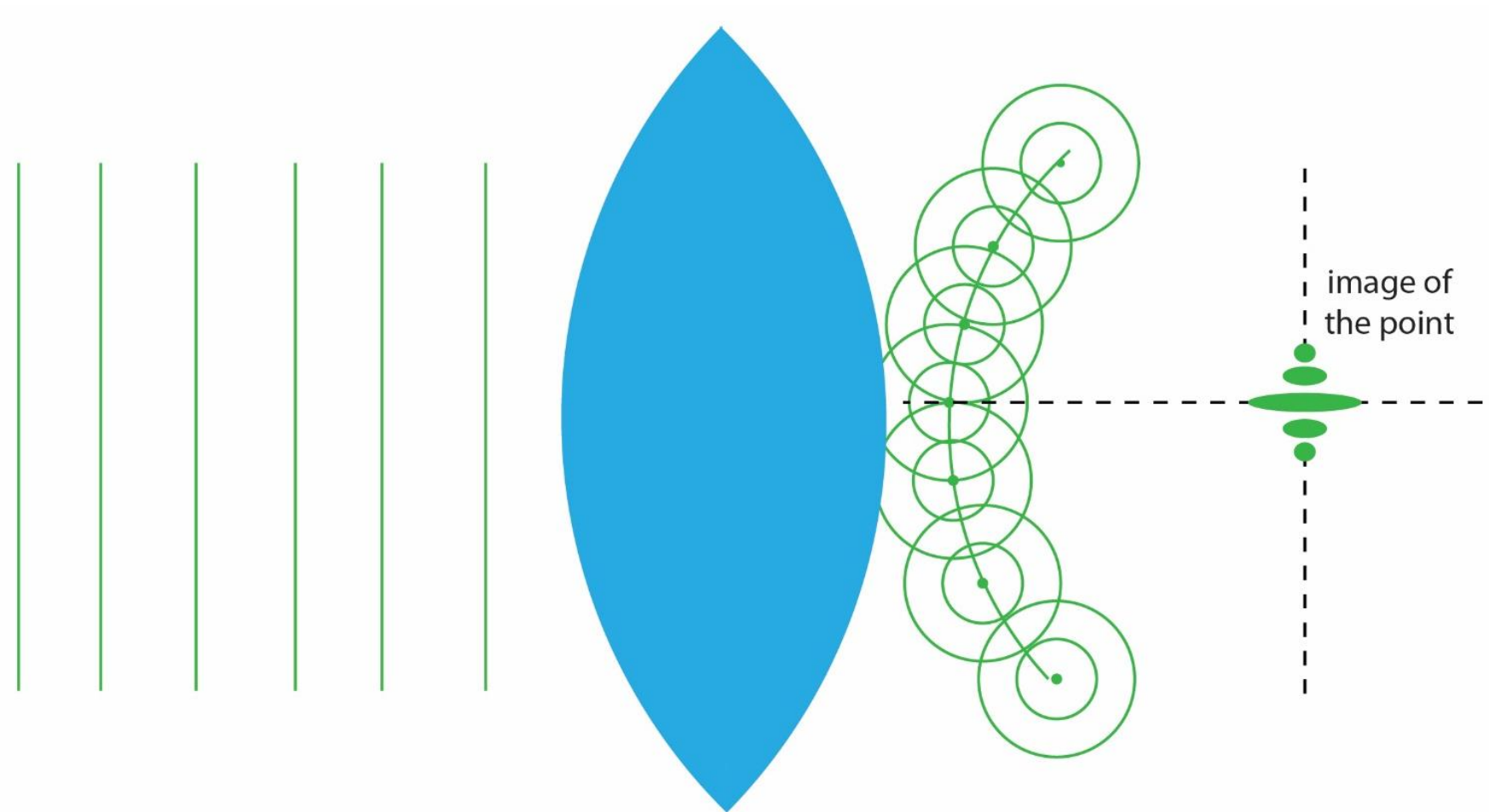
How the donut is created?



How the donut is created?



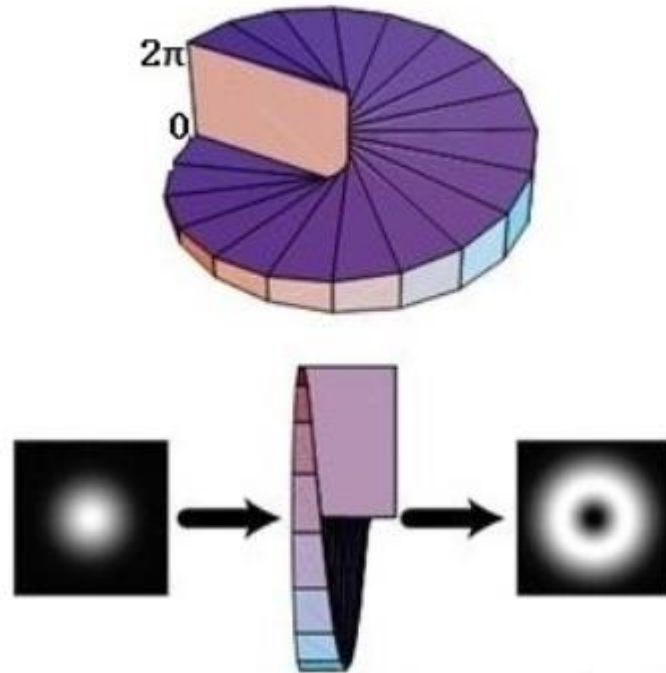
# How the donut is created?



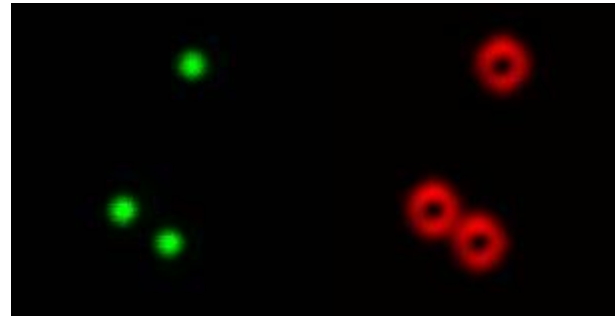


# The donut is created via optical vortex

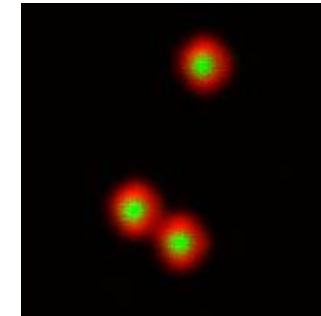
Courtesy of Courtial and O'Holleran, 2007



Excitation beam    Depletion beam



Overlay

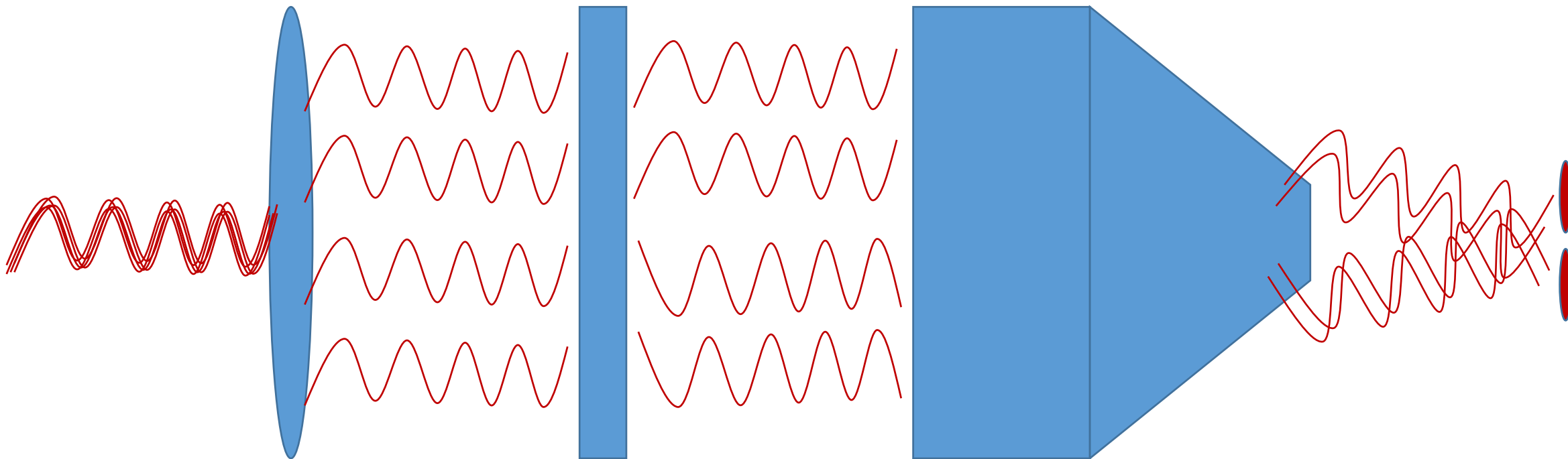


80 nm gold beads, 550 nm excitation, 660 nm depletion

Beam expander

Optical vortex

Objective

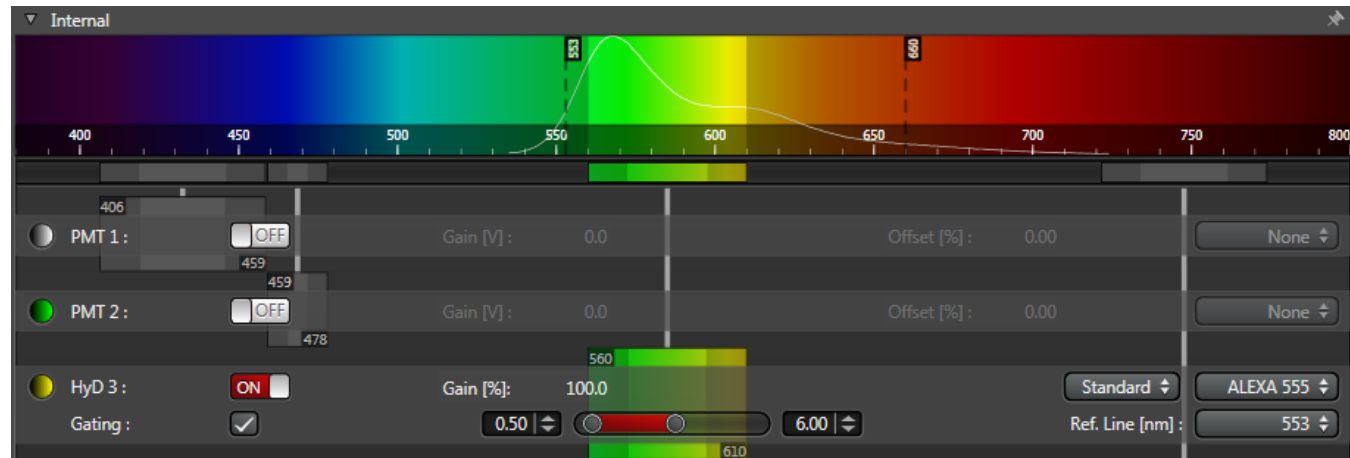


AF488

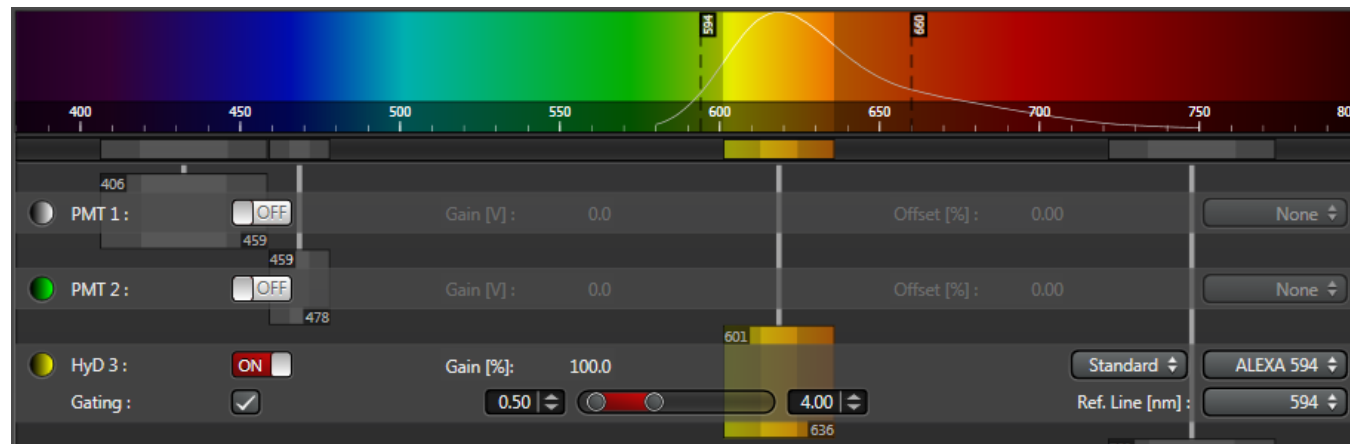


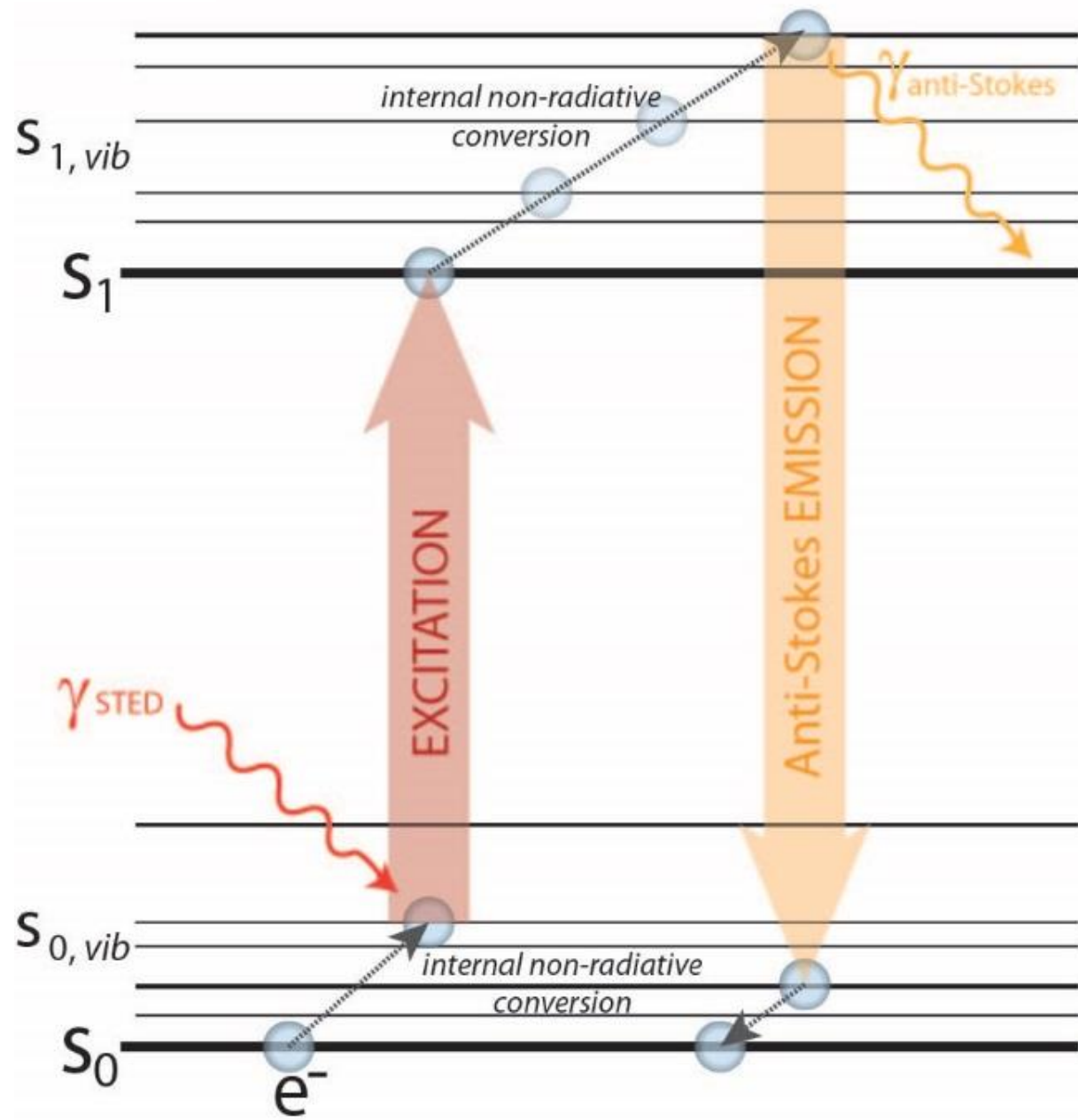
Depletion laser 660 nm

AF555

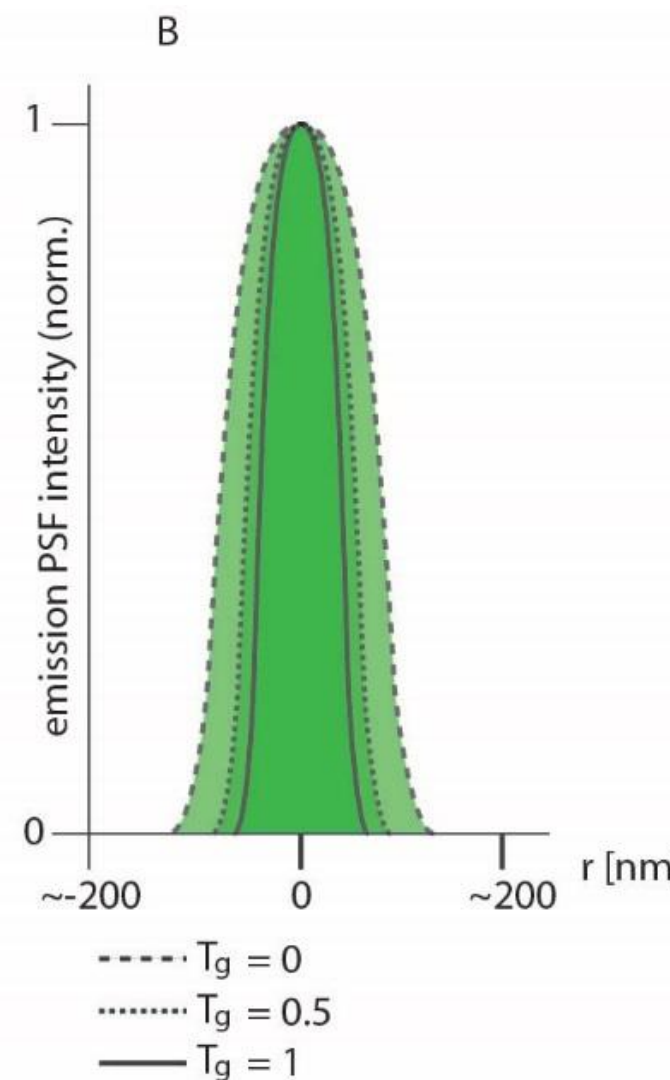
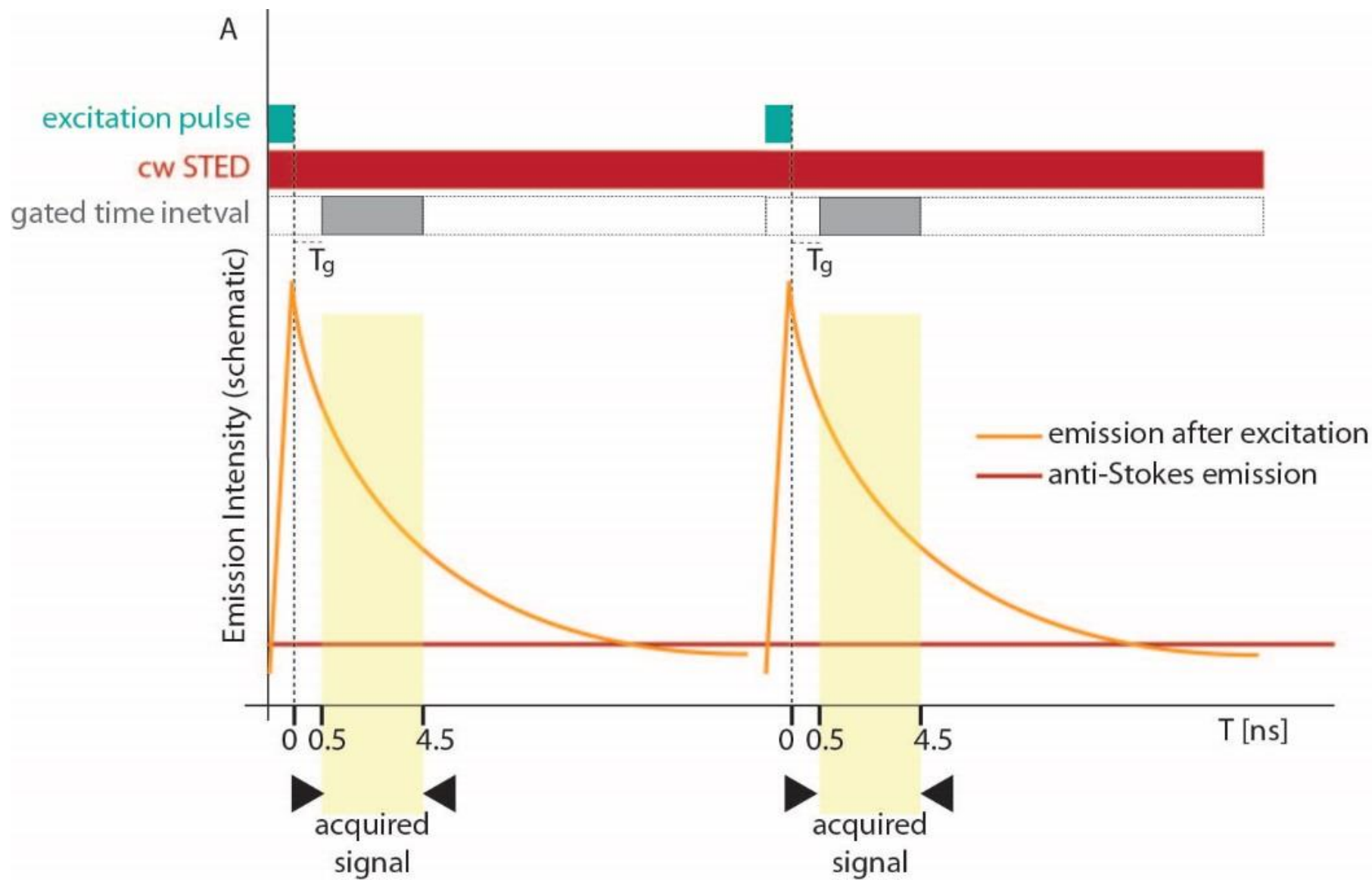


AF594

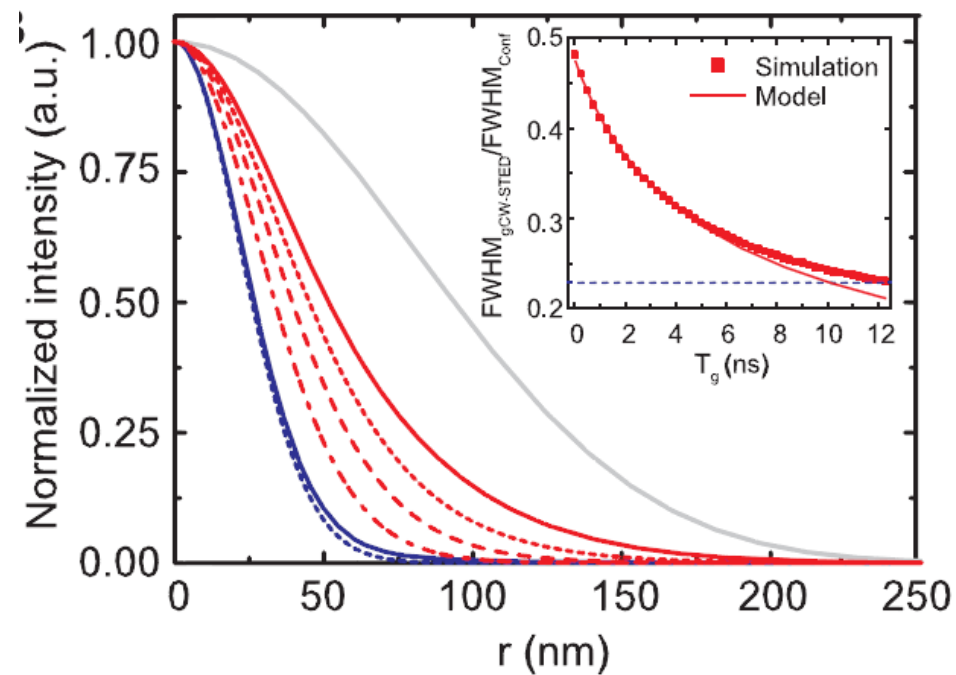
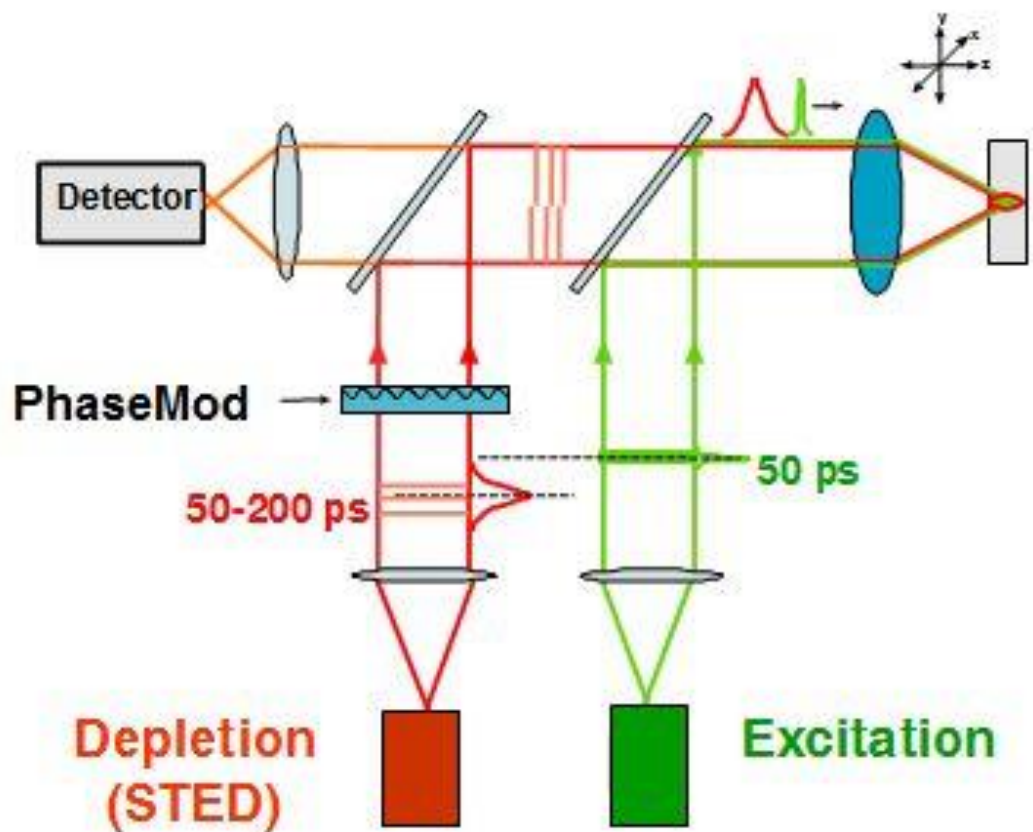




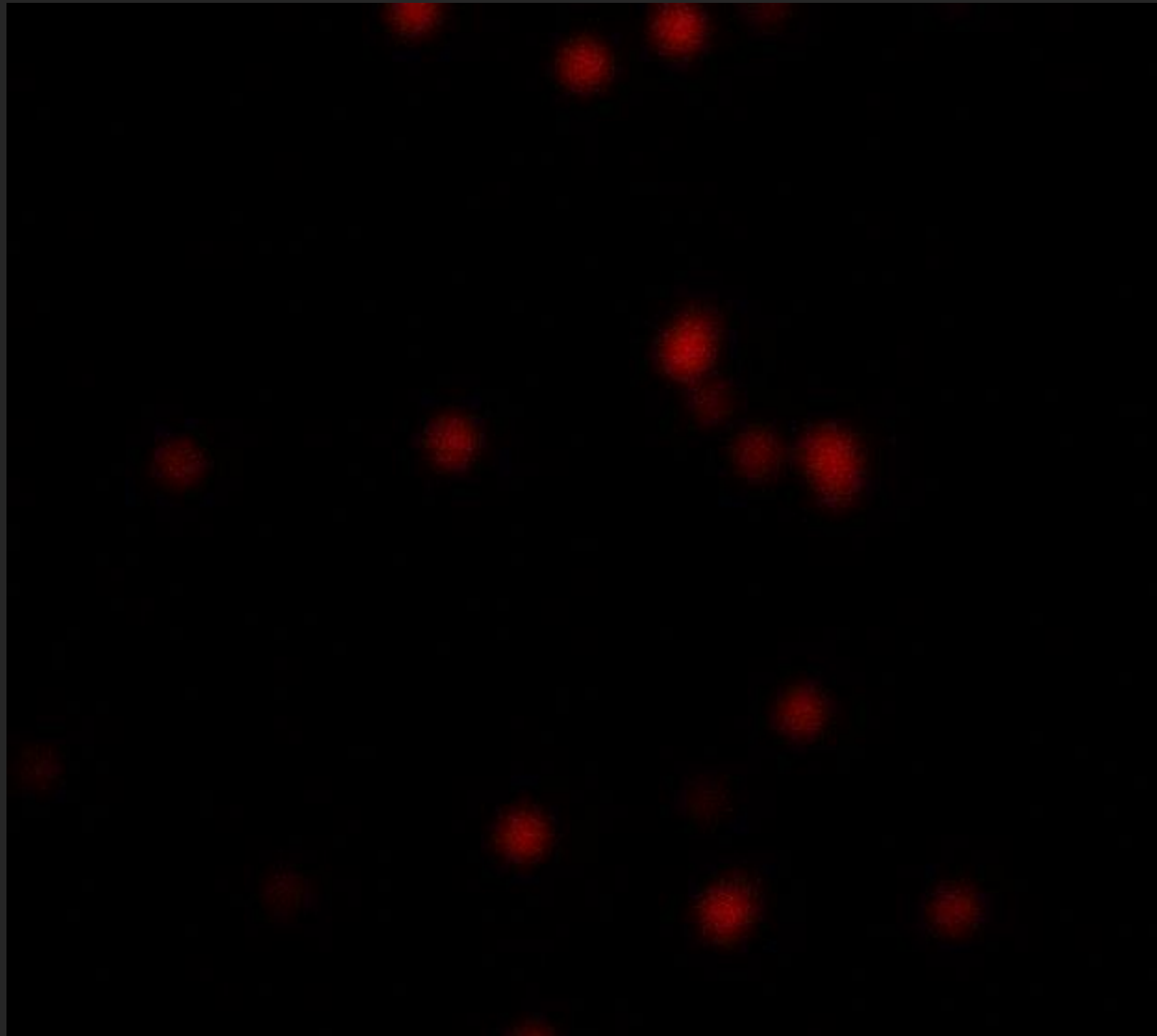
# Time Gating in continuous wave STED laser



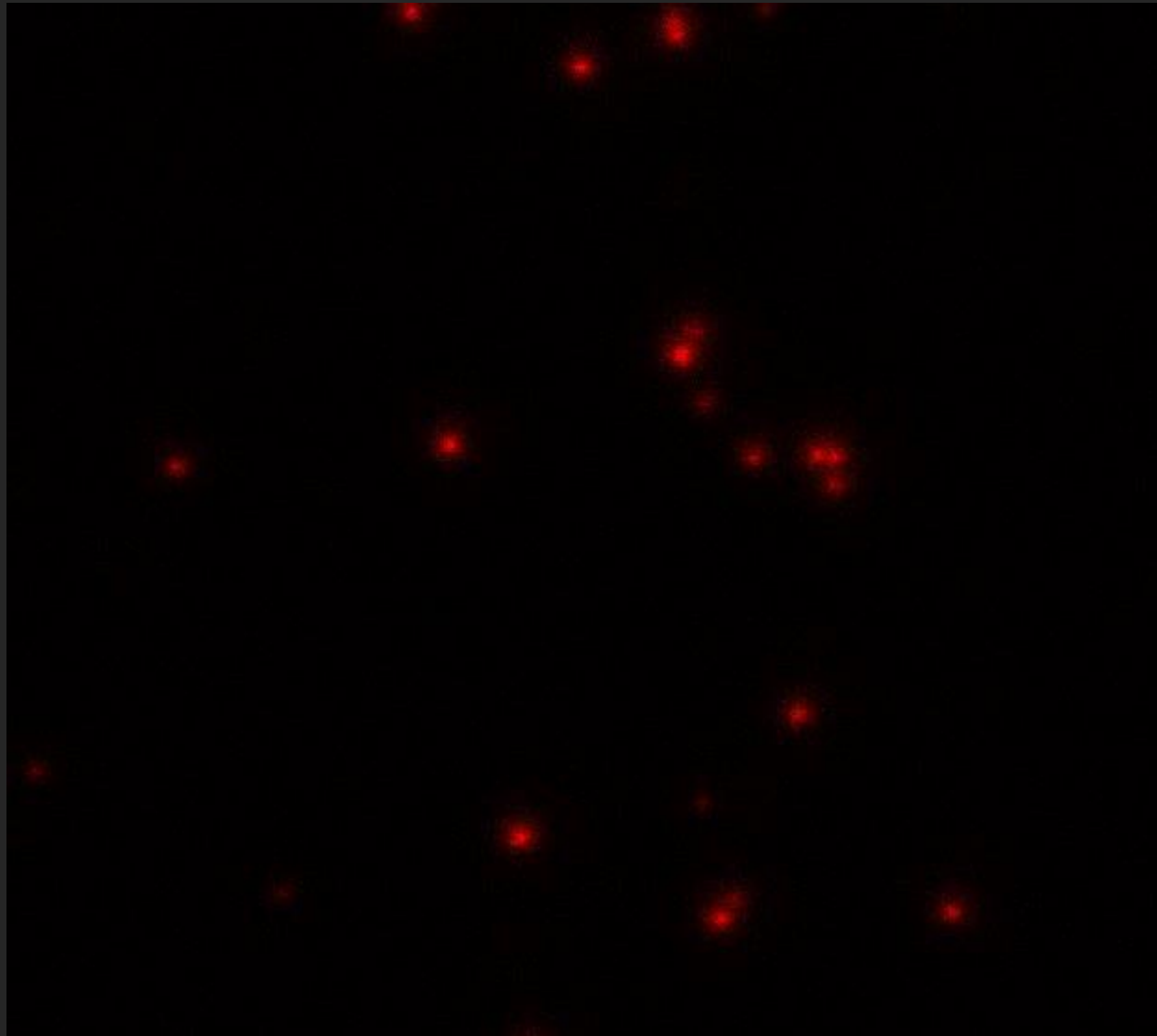
# Pulsed-STED and the time-gating in compare to CW-STED time-gating



48 nm beads; confocal

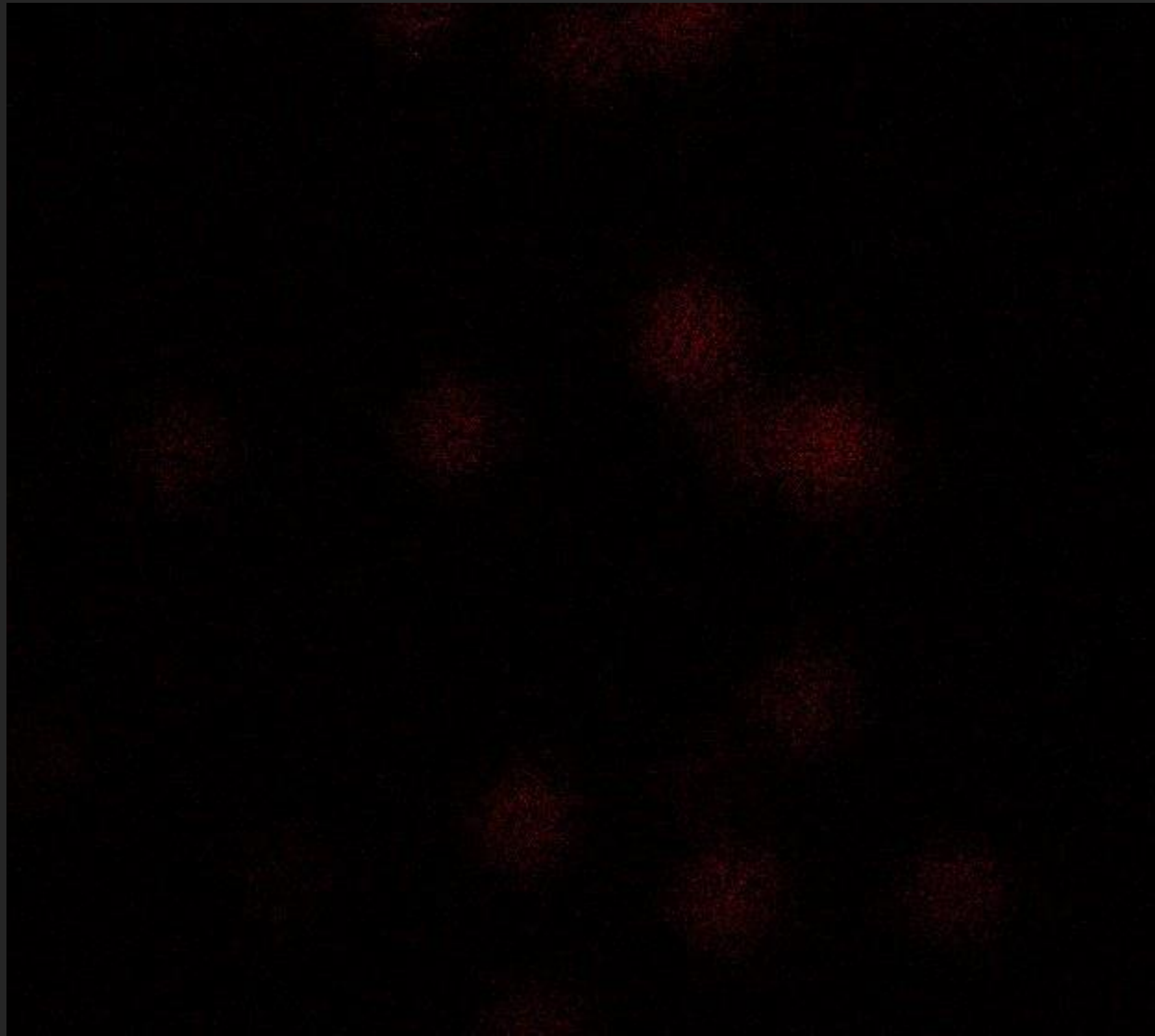


48 nm beads; non-gated cw-STED (660nm)





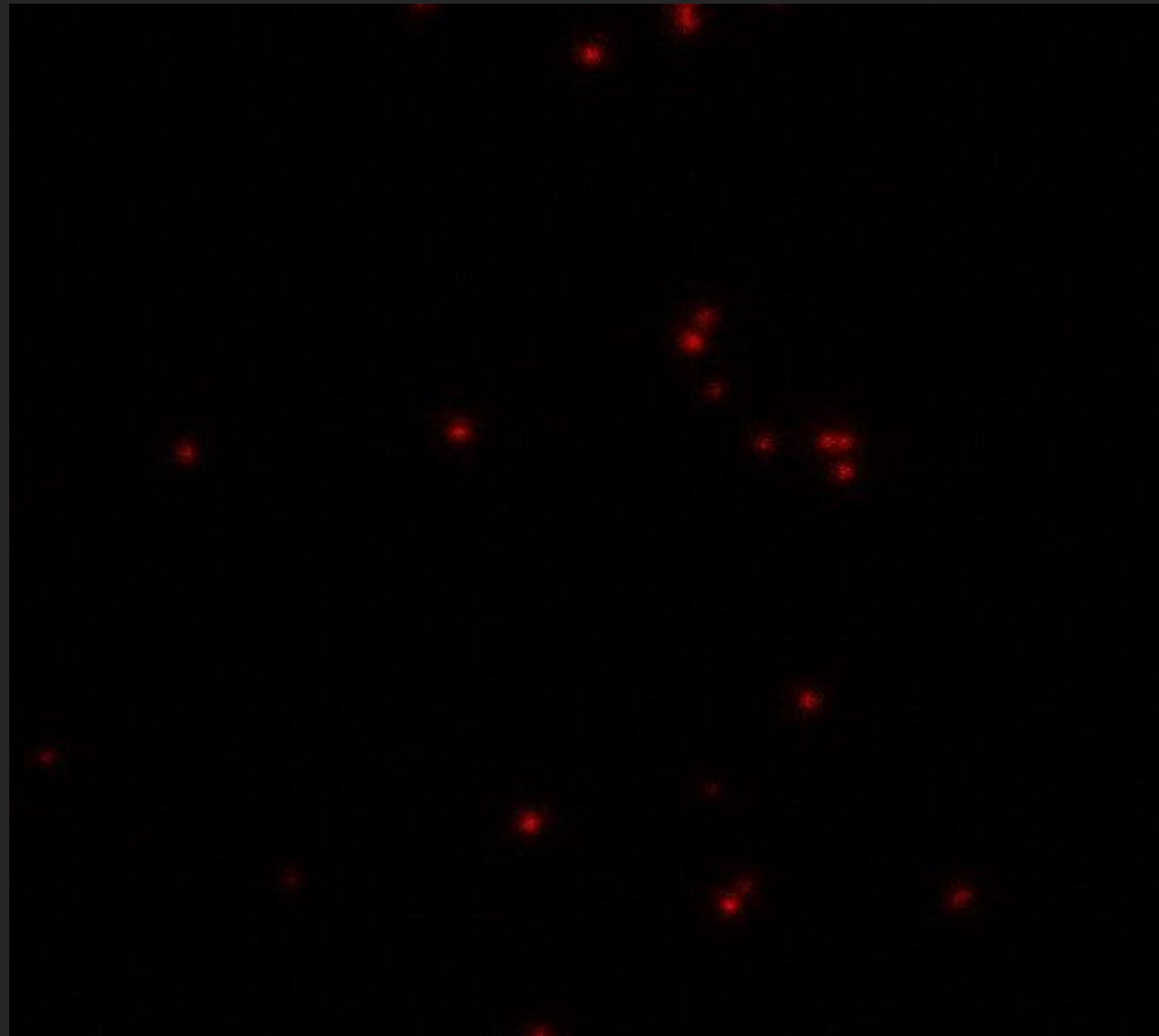
48 nm beads – anti-stokes signal, non-gated cw-STED (660nm)



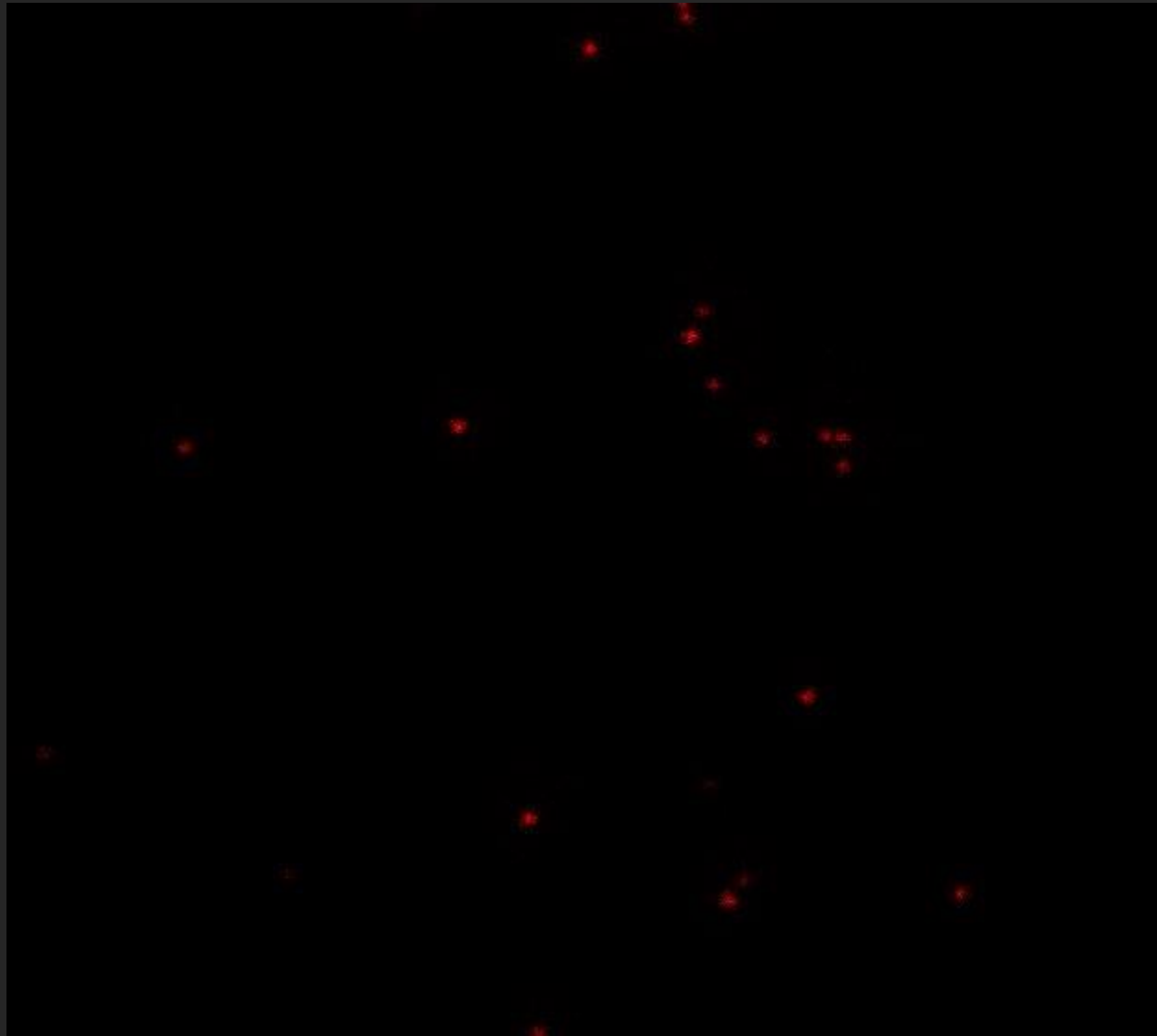
48 nm beads – anti-stokes signal, partially gated cw-STED (660nm)



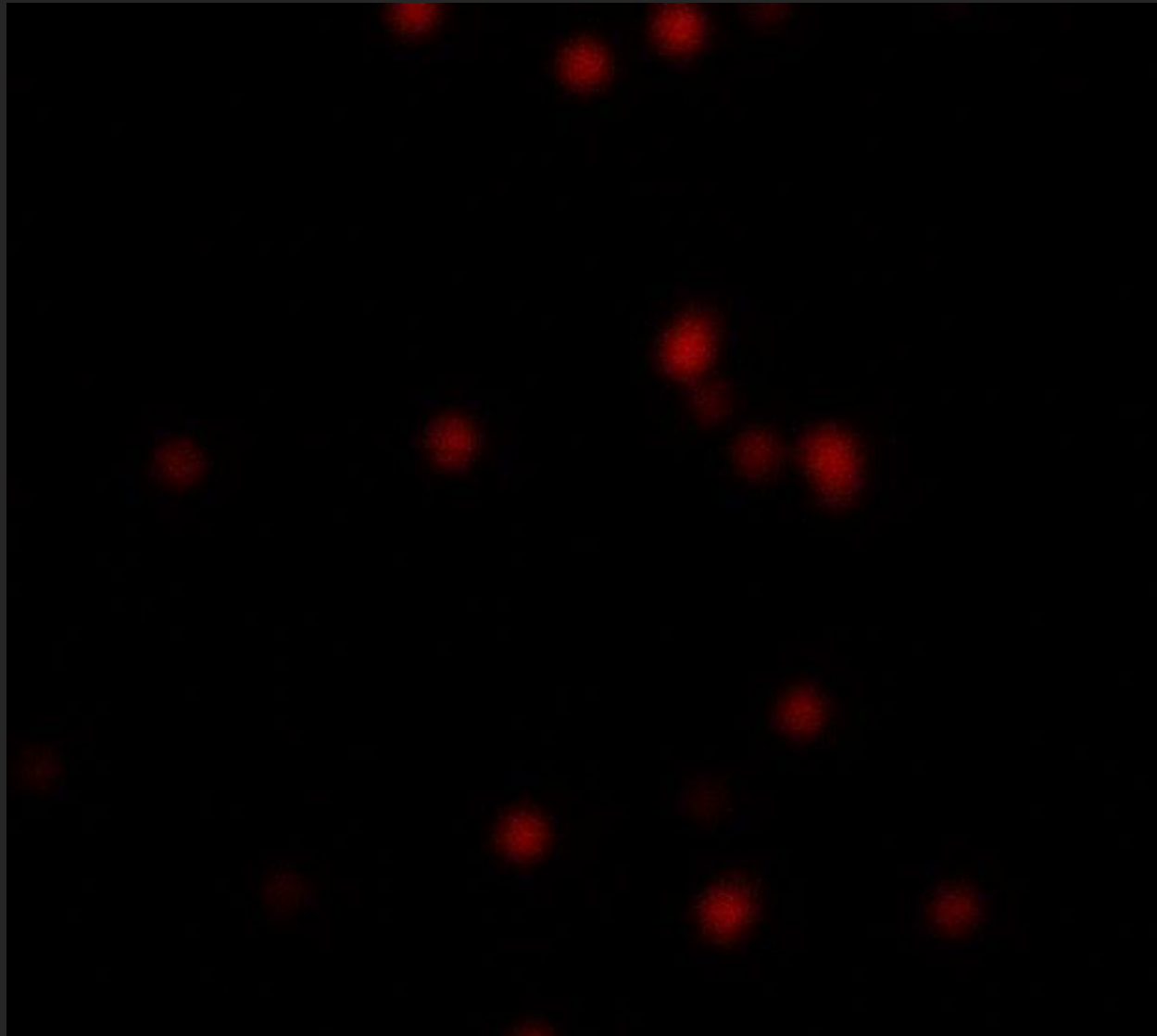
48 nm beads, partially gated cw-STED (660nm)



48 nm beads, gated cw-STED (660nm)



48 nm beads; confocal



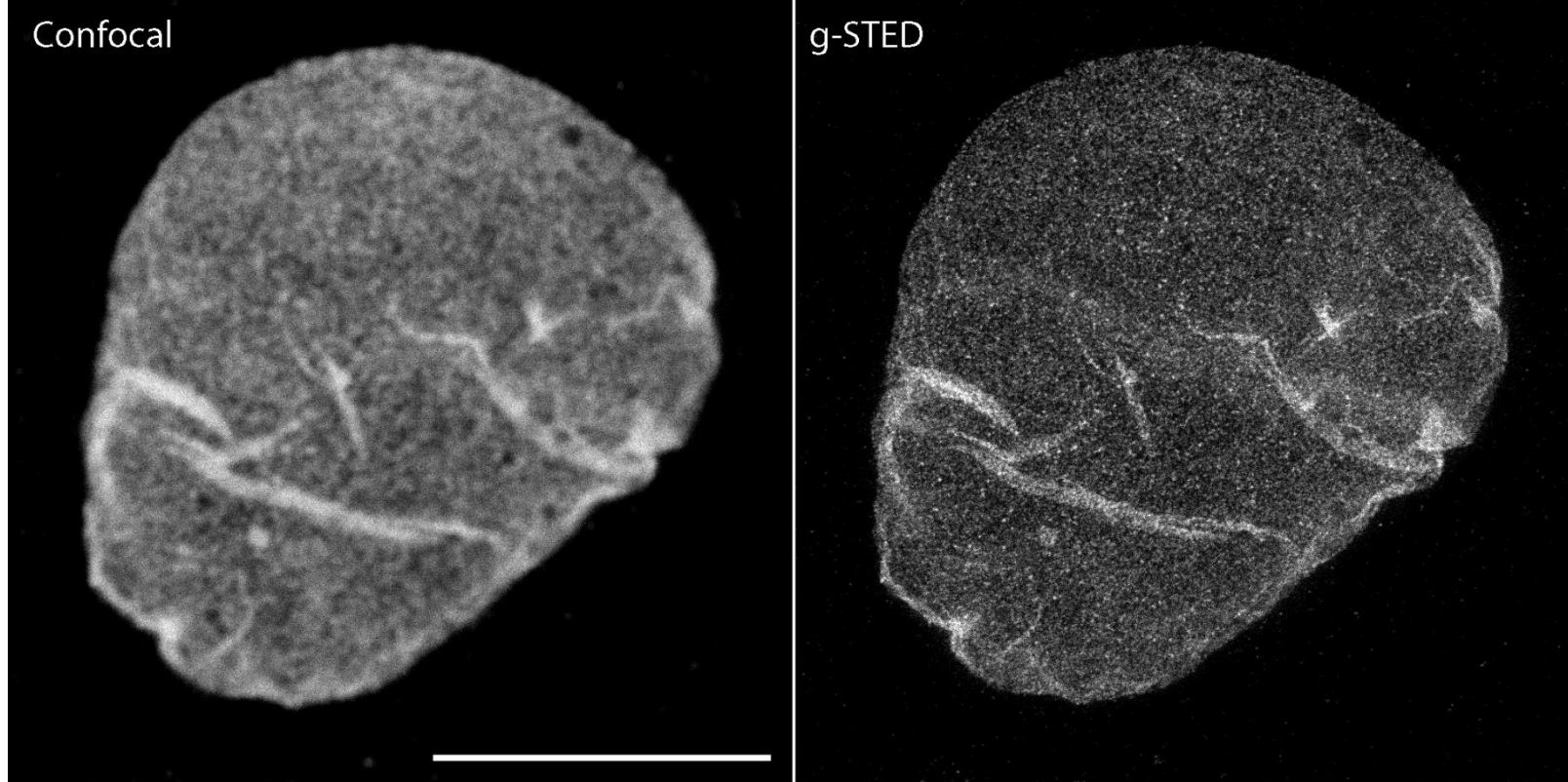
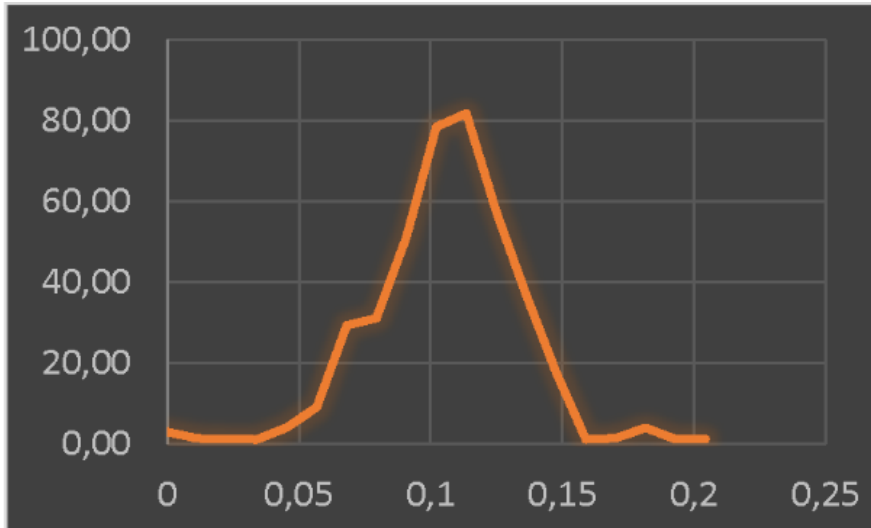
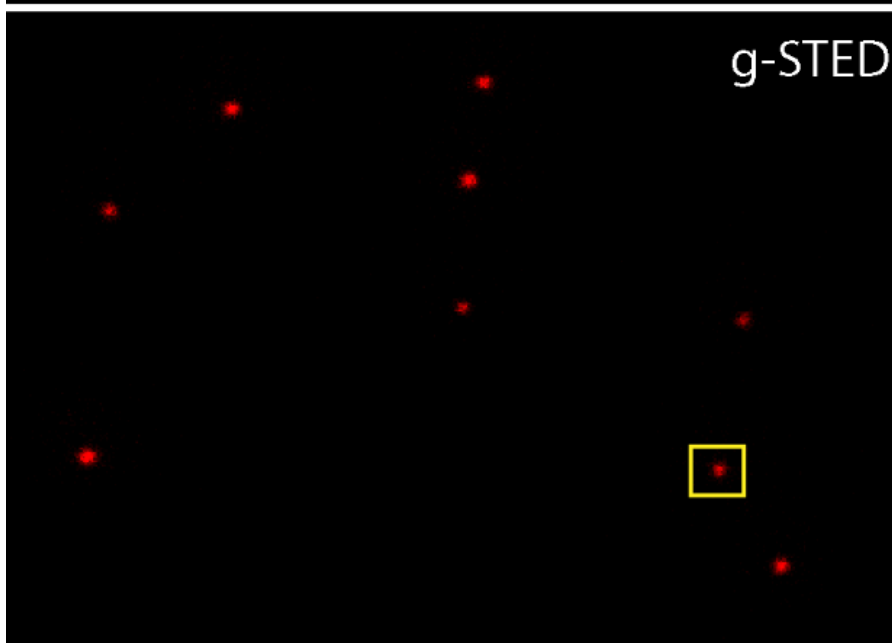
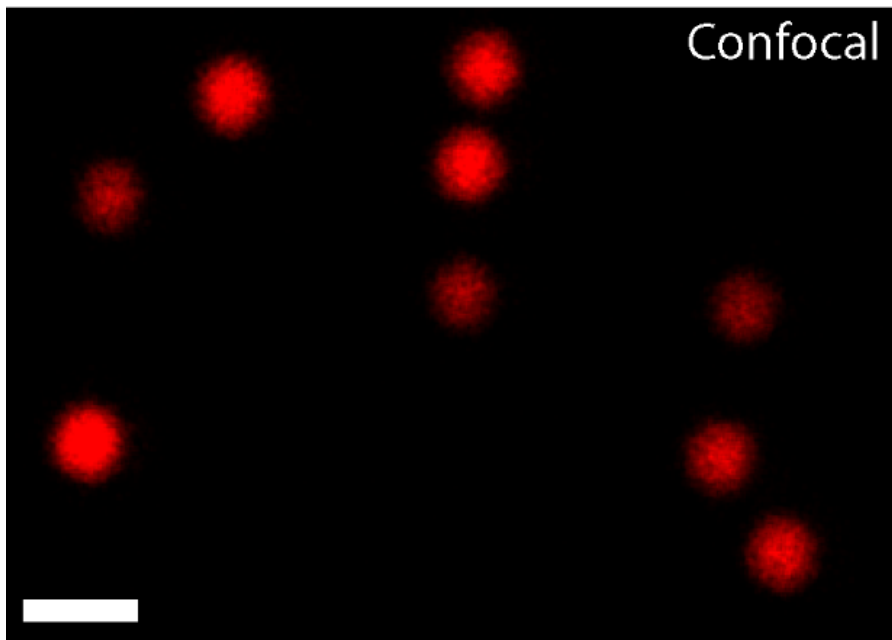


Image of U2OS cell nucleus; nuclear lamina visualized using indirect immunofluorescence, secondary antibody conjugated with AlexaFluor 555 (samples prepared in collaboration with *Jindriska Fiserova, Laboratory of Epigenetics of the Cell Nucleus, IMG CAS, Prague*)  
Images were acquired in Confocal and g-STED mode.

Leica TCS SP8 STED 3X  
660 nm depletion LASER  
Bar: 5  $\mu\text{m}$



Fluorescence bead sample optimized for STED, acquired in Confocal and g-STED mode (left top and bottom).

Plot profile (above) shows 50 nm FWHM. (X: microns, Y: intensities; 8bit depth)

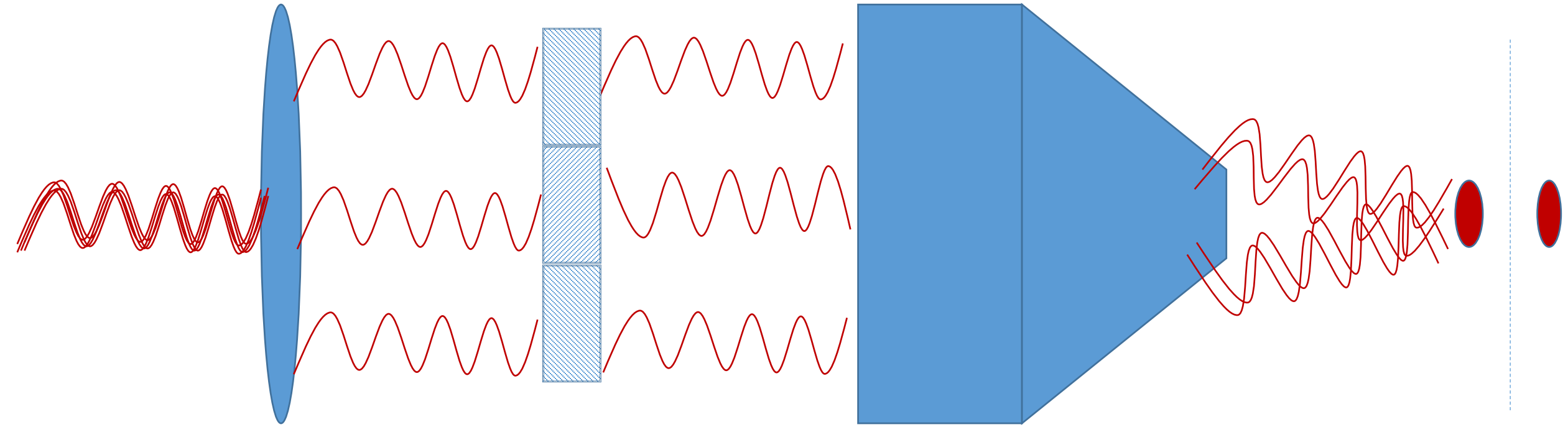
Leica TCS SP8 STED 3X  
660 nm depletion LASER  
Bar: 300 nm

# 3D STED

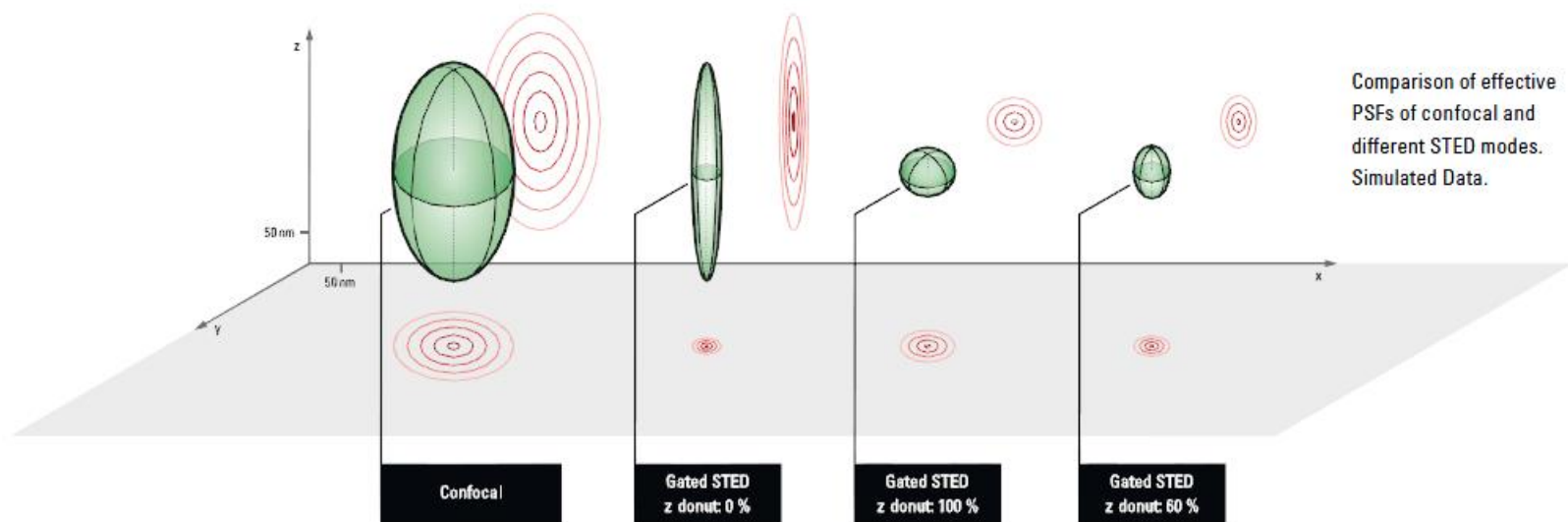
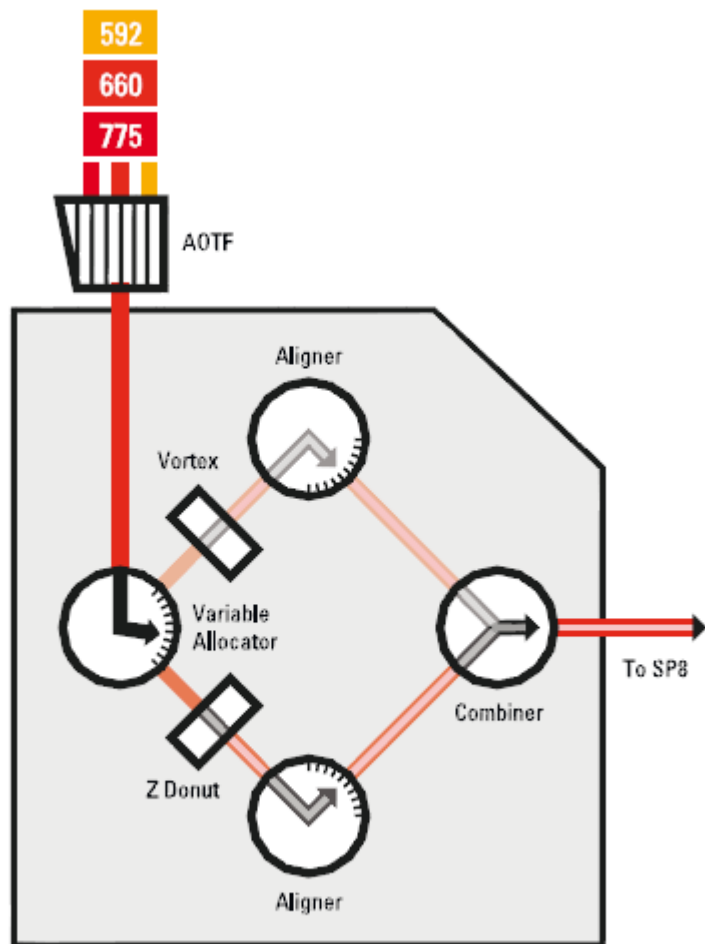
Beam expander

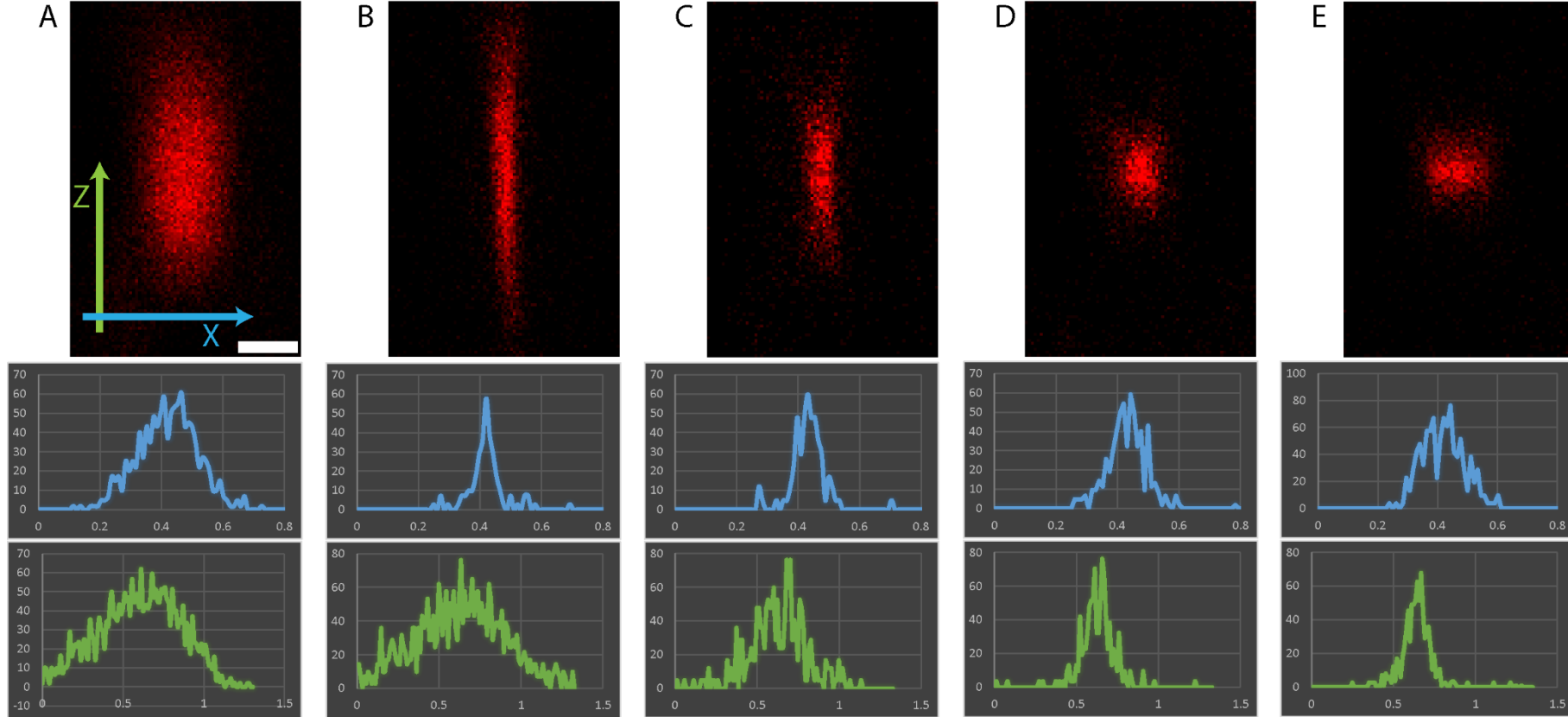
Z-Phase

Objective









PSF of fluorescence bead optimized for STED, image acquired using XZ scan.

Plotes above demonstrate the intensity profile of each PSF in X (blue) and Z (green) axis. The beads were imaged in the manner:

A: confocal

B: g-STED, 3D 0%

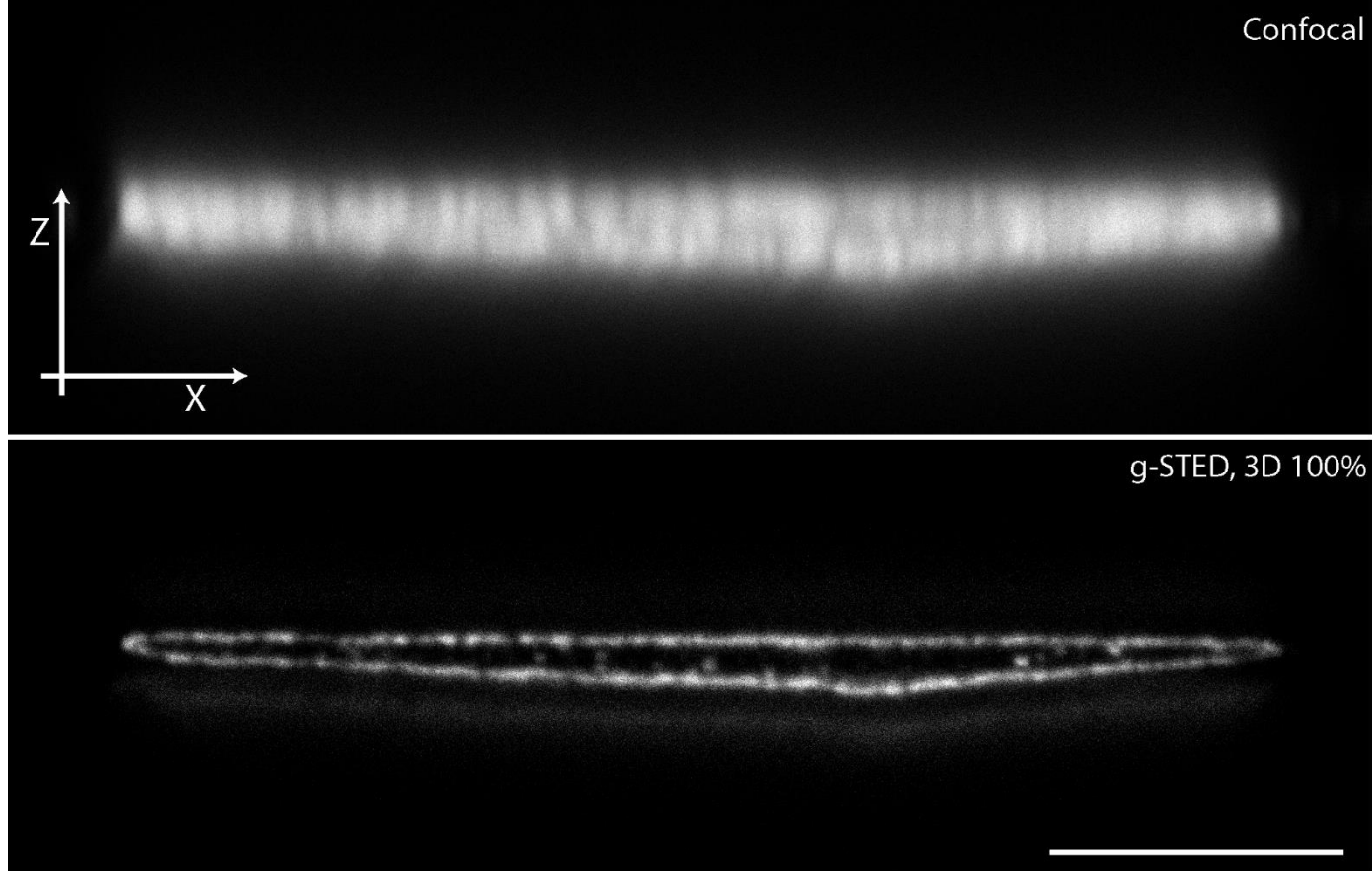
C: g-STED, 3D 20%

D: g-STED, 3D 70%

E: g-STED, 3D 100%

Leica TCS SP8 STED 3X  
660 nm depletion LASER

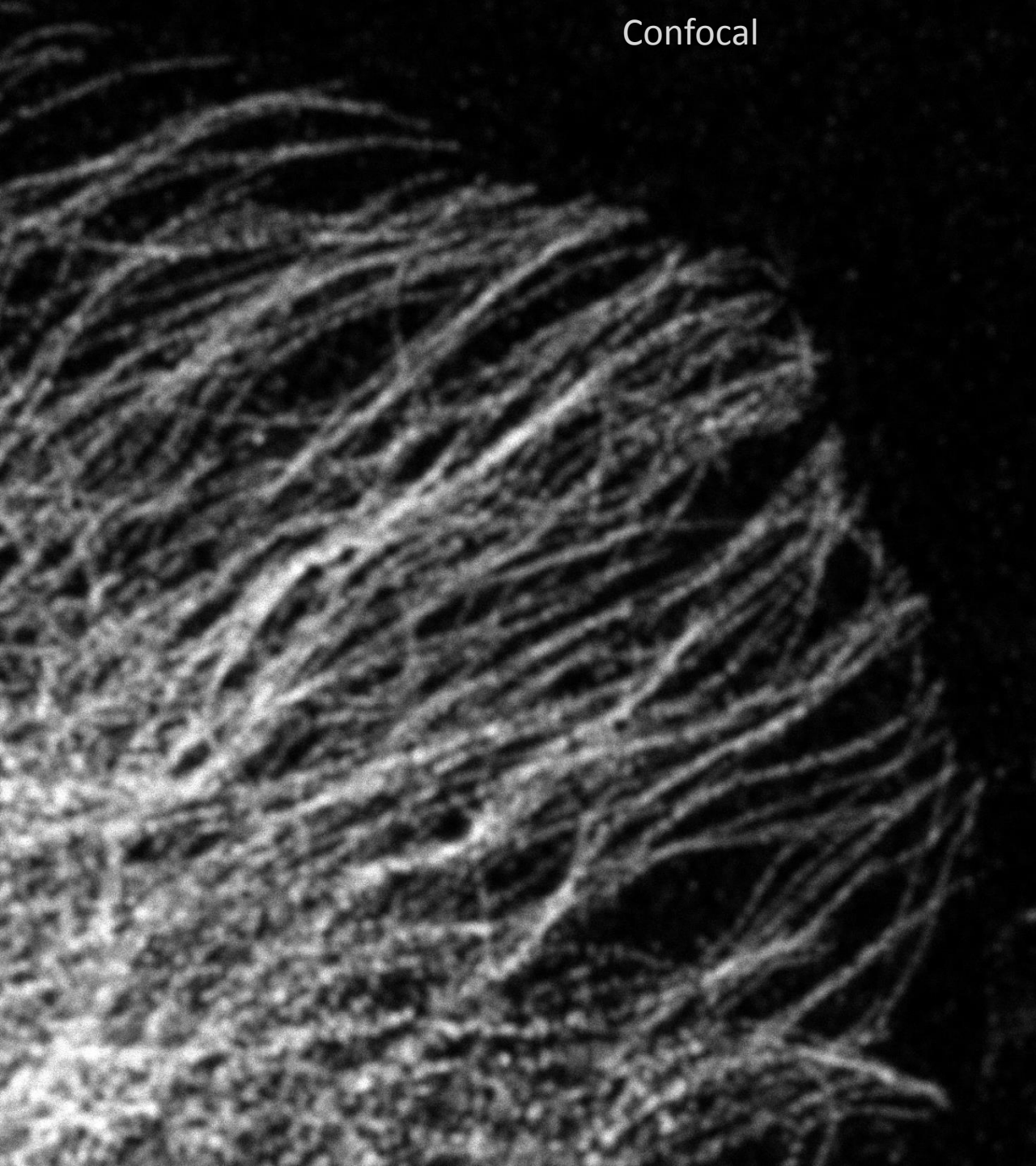
Bar: 300 nm



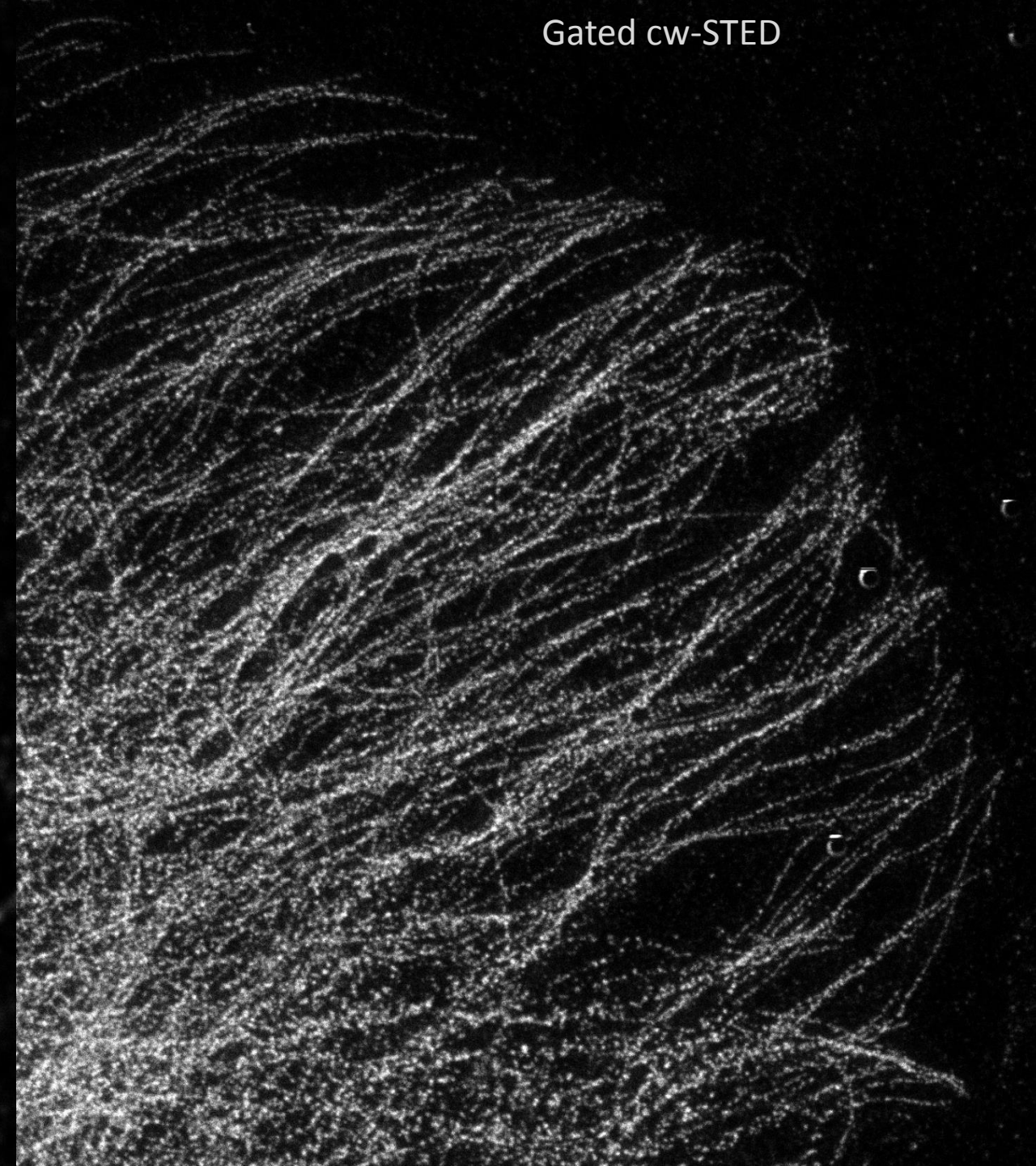
XZ profile of U2OS cell nucleus; nuclear lamina visualized using indirect immunofluorescence, secondary antibody conjugated with AlexaFluor 555 (samples prepared in collaboration with *Jindriska Fiserova, Laboratory of Epigenetics of the Cell Nucleus, IMG CAS, Prague*)  
Images were acquired using XZ scan in Confocal and g-STED mode.

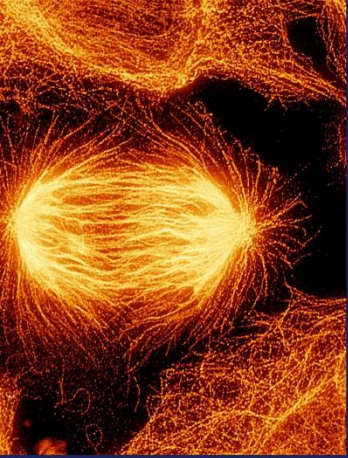
Leica TCS SP8 STED 3X  
660 nm depletion LASER  
Bar: 5  $\mu\text{m}$

Confocal



Gated cw-STED



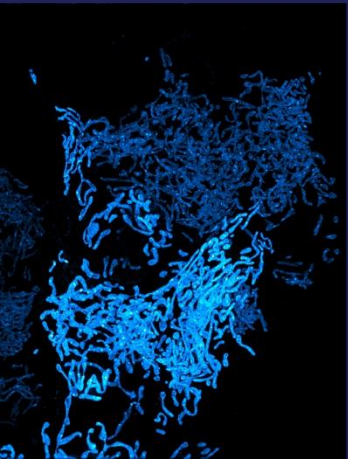
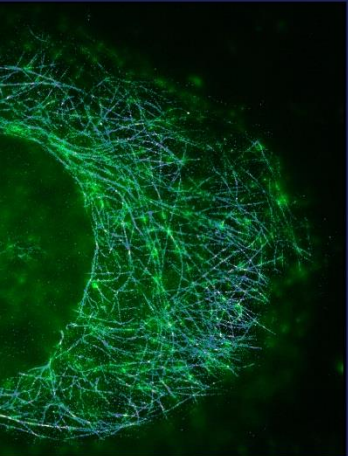


**Thank you for the attention!**

And many thanks to:

**Eduarda Dráberová, Klebanovych Anastasiya**  
Laboratory of Biology of Cytoskeleton, IMG

**Iva Jelínková**  
Laboratory of Biology of the Cell Nucleus, IMG



## Many thanks for support:

Institute of Molecular Genetics, CAS

Czech-Bioimaging Infrastructure – LM2015062

“Centre of Model Organisms” OPVK (CZ.2.16/3.1.00/21547)

“Biomodels for health” (LO1419)

CZ.02.1.01/0.0/0.0/16\_013/0001775 Modernizace a podpora výzkumných aktivit národní infrastruktury pro biologické a medicínské zobrazování Czech-BioImaging

